ISSN: 2319-7692 (Print) ISSN: 2319-7706 (Online)

IJCMAS

INTERNATIONAL JOURNAL OF CURRENT MICROBIOLOGY AND APPLIED SCIENCES



WWW.IJCMAS.COM

INDEX





Original Research Articles



PRINT ISSN : 2319-7692 Online ISSN : 2319-7706 Issues : 12 per year Publisher : <u>Excellent Publishers</u> Email : editorijcmas@gmail.com / submit@ijcmas.com

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 432-441 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.050</u> Implementation Probiotics Cellulolitic B-7 Bacteria (Isolation from Buffalo Rumen) into Rations on the Performance, Abdominal Fat and Serum Cholesterol of Duck Eny Puspani*, Desak Putu Mas Ari Candrawati and Dan I.G.N.G. Bidura Faculty of Animal Science, Udayana University, Denpasar, Bali, Indonesia Jl. PB. Soedirman, Denpasar, Bali-Indonesia

**Corresponding author*

Abstract:

This research aims to study the influence of cellulolitic B-7 bacterial isolates rumen of buffalo as a source of probiotics on the performances, abdominal fat, and cholesterol serum of a male Bali duck. A total of 240 male bali ducks were randomized in a completely randomized design (CRD) with four treatments and six replications. Treatment is attempted is the ducks that were given rations without supplementation of probiotic cultures cellulolitic as control (A); ration with 0.20% (B); 0.40%; and 0.60% probiotic cultures selulolityc B-7, resvectively. The isolate has passed the test and has been considered as potential probiotic according to our previous study (Bidura *et al.*, 2014). The results showed that the ducks were given rations containing probiotics result in weight gain and feed efficiency significantly (P<0.05) better than the control. However, administration of probiotics through diet significantly (P<0.05) can reduce abdominal fat weight and serum kolseterol in ducks. From these results it can be concluded that supplementation of 0.20% to 0.60% probiotic cellulolitic B-7 bacteria were isolated from the rumen of buffalo in the ration can significantly improve live weight gains and feed efficiency of Bali duck. In contrast, significantly reduced abdominal-fat weight and serum cholesterol of ducks.

Keywords: Probiotics, buffalo rumen, performance, cholesterol, abdominal-fat, duck.

Download this article as

How to cite this article:

Eny Puspani, Desak Putu Mas Ari Candrawati and Dan I.G.N.G. Bidura. 2016. Implementation Probiotics Cellulolitic B-7 Bacteria (Isolation from Buffalo Rumen) into Rations on the Performance, Abdominal Fat and Serum Cholesterol of Duck. *Int.J.Curr.Microbiol.App.Sci.* 5(11): 432-441 doi:<u>http://dx.doi.org/10.20546/ijcmas.2016.511.050</u>

Editorial Board

Editor-in-Chief

Dr.M.Prakash,M.Sc.,M.Phil.,Ph.D., Head, Department of Microbiology Kancheepuram, Tamilnadu, India.

Executive-Editors

Dr. Ravish Kumar Chauhan Department of Chemistry, Indira Gandhi National

College,Ladwa (Kurukshetra)

Haryana, India

Dr.M.H.Fulekar, Professor of Environmental Biotechnology

Department of Life Sciences, University of Mumbai Santacruz (E) Mumbai, India

Dr. Subbiahpoopathi

Scientist-F Vector Control Research Centre (Ministry of Health & Family Welfare, Govt.of India) Medical Complex, Indira Nagar,Pondicherry, INDIA

Dr. K.Madhava Chetty,

Plant Taxonomist, Department of Botany, SVUCS, S.V. University, Tirupati, India

Dr.N.Sivakumar, M.Sc., Ph.D.,

Assistant Professor, Sultan Qaboos University Muscat, Oman.

Editorial Advisory Board

- 1) Dr.N.Karmegam, Govt. arts and Science, Salem, India
- 2) Dr. Thilagavathy Daniel, The Gandhigram Rural Institute-Deemed University, India.
- 3) Dr. B. Kadalmani, Bharathidasan University, India.
- 4) Dr. S. Karuppusamy, Madura College, India.
- 5) Dr. Arvind Bijalwan, Faculty of Technical Forestry, Indian Institute of Forest Management (IIFM), An Autonomous Institute of Ministry of Environment, Forests & Climate Change, Government of India, Bhopal, Madhya Pradesh, India
- Dr.C.M.Ramakritinan, Department of Marine and Coastal Studies, School of Energy, Environment and Natural Resources Madurai Kamaraj University, Madurai – 625 021, Tamilnadu
- 7) Dr.T.Sathishkumar, MCC, Chennai, India
- 8) Dr.A.M.Saravanan, Caledonian University, Oman
- 9) Dr.A.Arunachalam, King Saud University, UAE
- 10) Dr.Zhengchao Wang, Provincial Key Lab for Developmental Biology and Neuroscience College of Life Science (Qishan Campus), Fujian Normal University, P. R. China

- 11) Dr. Shahzad Akbar Khan (I.S), Department of Pathobiology Faculty of Veterinary and Animal Sciences, The University of Poonch, Rawalakot, Pakistan
- 12) Dr.Hazim Jabbar Shah Ali Al-Daraji, Animal Resource Department, College of Agriculture, University of Baghdad, Abu-Ghraib, Iraq
- 13) Dr. Subhash Chand, Division of Soil Science, Faculty of Agriculture, Wadura, Sopore-193201, Jammu and Kashmir, India
- 14) Dr.Maged El-ashker, Department of Internal Medicine and Infectious Diseases, Faculty of Veterinary Medicine Mansoura University, Egypt
- 15) Dr.M.Jayakumar, Assistant Professor, Department of Chemical Engineering, Institute of Technology, Haramaya University, Dire Dawa, Ethiopia
- 16) Dr. N. Amaresan, C G Bhakta Institute of Biotechnology, Gujarat, India
- 17) Dr. Sushama Talegaonkar, Department of Pharmaceutics, Faculty of Pharmacy, Jamia Hamdard, New Delhi, India
- 18) Dr.B.Ramesh, Sri Sankara College of Arts and Science, Enathur, India
- 19) Dr.Ugwu Okechukwu Paul-Chima, Department of Biochemistry, University of Nigeria Nsukka, Enugu State, Nigeria.
- 20) Dr. Elvis-Enowbeyang Tarkang, South West Region, Republic of Cameroon
- 21) Dr.S.M.Gopinath. Head, Department of Biotechnology, Acharya Institute of Technology, Bangalore, India
- 22) Dr.Philippe Sessou, Cotonou / Benin
- 23) Dr.Satyanand Tyagi, President & Founder, Tyagi Pharmacy Association(TPA) & Scientific Writer (Pharmacy), New Delhi, India-110074.
- 24) Dr. Soleiman Mahjoub, Ph.D.Professor of Clinical Biochemistry, Babol University of Medical Sciences, Babol, Iran.
- 25) Dr. Shamim Ahamad, Aligarh Muslim University, Aligarh, UP, India
- 26) Dr.S.Vijayanand, Thiruvalluvar University, India
- 27) Dr. Parwez Qureshi, College of Science (Girls), Jazan University, Jazan Kingdom of Saudi Arabia
- 28) Dr. Subha Ganguly, Faculty of Fishery Sciences, West Bengal University of Animal and Fishery Sciences, Kolkata, India
- 29) H.A.Sayeswara, Department of Zoology, Sahyadri Science College(Autonomous), Shivamogga-577203, Karnataka state
- 30) Dr.Sami R. Al-Zubaydi, Biology Department, Faculty of Science. Duhok University, Kurdistan Region of Iraq.
- 31) Dr.P.J Hisalkar, Department of Biochemistry, People"s College of Medical Sciences & Research Centre, Karond Bypass Road, Bhanpur, Bhopal, Madhya Pradesh, India
- 32) Dr. Mousavi Khaneghah Amin, Brazil St- Vanak Sq- Tehran- Iran.
- 33) Dr.Anand Prem Rajan, School of Bio Sciences and Technology, Vellore Institute of Technology University, Vellore, India
 35) Dr.Sankhadip Bose, Department of Pharmacognosy, Gupta College of Technological Sciences, College of Pharmacy, Ashram More, G. T. Road, Asansol, West Bengal, India
- 34) Dr.Sonali Jain, Department of Microbiology, Goldfield institute of medical sciences, Faridabad, India
- 35) Dr. D. Sathish Sekar, Department of Biotechnology, Arignar Anna College (Arts & Science) Jakkappan Nagar, Krishnagiri, Tamilnadu, India
 38) Dr.Tara Chand, Professor & Principal at Regional College of Pharmacy, Jaipur Rajastan, India

36) Dr.A.R.Gulnaz, Department of Biochemistry at Farooqia Dental College &Hospital, Mysore, India

40) Dr.Ahmet ADIGUZEL, Department of Molecular Biology and Genetics, Faculty of Science, Ataturk University, Turkey

- 37) Dr. Vinay B. Raghavendra, Department of Biotechnology, Teresian College, Affiliated to University of Mysore, India
- 38) Dr. Arvind Bijalwan, Faculty of Technical Forestry, Indian Institute of Forest Management (IIFM), Ministy of Environment and Forests, Government of India, Nehru Nagar, Bhopal–, Madhya Pradesh, India
- 39) Dr. Shivaji P. Chavan, Associate Professor, Department of Zoology, School of Life Sciences, Swami Ramanand Teerth Marathwada University, Nanded- 431606, Maharashtra, India.
- 40) Dr. Ruwan Duminda Jayasinghe, Department of Oral Medicine and Periodontology, Faculty of Dental Sciences, University of Peradeniya, Peradeniya, Sri Lanka
- 41) Dr. P. Pandia Vadivu, Division of Cognitive Science, School of Education, Tamilnadu Open University, Chennai, India
- 42) Dr.Sanjay Shamrao Nanware, Research and Post Graduate Department of Zoology, Yeshwant Mahavidyalaya, Nanded M.S, India
- 43) Dr.Nagham Mahmood Aljamali, Department of Chemistry, IRAQ
- 44) Dr.Saher Mahmood Jwad Aljamali, Biology Department Kufa University Iraq
- 45) Dr.Rasha Khalil Al-Saad, Veterinary Medicine, Iraq
- 46) Dr.Rachana Bhatt, Department of Biomedical Engineering, New Jersey Institute of Technology, NJ, USA
- 47) Dr.Said Elshahat Abdallah, Department of Agricultural Engineering, Faculty of Agriculture, Kafrelsheikh University, Kafr Elsheikh, Egypt
- 48) Dr. D. Dhanavel, Department of Botany, Annamalai University, Annamalainagar, INDIA
- 49) Dr.Swaminathan Palanisami, Center for Bioenergy, Cooperative Research, Lincoln University of Missouri Jefferson City, Missouri 65101
- 50) Ling-juan Zhang, Laboratory of Richard Gallo, Division of Dermatology, School of Medicine, University of California, San Diego, CA 92121, USA 55) Dr. Abd El-aleem Saad Soliman Desoky, Department of Plant Protection, Faculty of Agriculture, Sohag Univ., Sohag, Egypt
- 51) Dr. Idress Hamad Attitalla, Department of Microbiology (Head), Faculty of Science, Omar Al-Mukhatr University, Al-Bayda, Libya
- 52) Dr.C.Manikandan, Assistant Professor, P.G & Research Dept. of Microbiology, Marudupandiyar College (Arts & science), Thanjavur, Tamilnadu, India
- 53) Dr.S.Prakash, Professor and Director Research, Nehru Institute of engineering and Technology, Nehru Group of Institutions, Coimbatore, India
- 54) Dr. Khaled Abdel Rahman Youssif Elbanna, Faculty of Agriculture, Department of Agricultural Microbiology Fayoum University, Fayoum, Egypt
- 55) Dr. S. Umavathi, PG and Research Department of Zoology J.K.K. Nataraja College of Arts & Science Komarapalayam 638183, India
- 56) Dr. Jon Ver Halen, Department of Surgery, Texas A&M University School of Medicine, Texas
- 57) Dr. Sashikala Beura, Biotechnology-cum-Tissue Culture Centre, Orissa University of Agriculture and Technology (OUAT), Bhubaneswar, India

- 58) Dr. Mohamed Ali Mohamed Eldeeb, University of Alberta, Edmonton, Alberta, Canada
- 59) Dr. C. Viswanatha, Arba Minch University, Ethiopia
- 60) Dr. Hany A. Fouad, Faculty of Agriculture, Sohag University, Sohag, Egypt66) Dr. Atul Bhargava, Amity Institute of Biotechnology, Amity University Lucknow Campus, Lucknow, India
- 61) Dr. A. Heidari, California South University (CSU), Irvine, California, USA
- 62) Dr. Hema Chandra Kotamarthi, Massachusetts Institute of Technology, Cambridge, USA
- 63) Dr. Moustafa Mohamed Sabry Bakry, Plant Protection Res. Inst., Agric. Res. Center, Giza, Egypt
- 64) Dr. Ram Swaroop Meena, Assistant Professor, Deapartment of Agronomy, Inistitute of Agricultural Sciences, BHU, Varanasi (UP)- 221 005, India
- 65) Dr. Refat A. Youssef, Professor of plant nutrition and soil fertility, Soils and Water Use Dept., National Research Centre Dokki, Giza, Egypt
- 66) Dr.Javad Sharifi-Rad, Department of Pharmacognosy, Faculty of Pharmacy, Zabol University of Medical Sciences, Zabol, Iran
- 67) Dr. Hammad Ahmad Khan, Associate Professor, Department of Zoology, Wildlife and Fisheries, Faculty of Science, University of Agriculture, Faisalabad
- 68) Dr. YahyaJirjees Salman Hamied, College of Medicine, Kirkuk University, Iraq
- 69) Dr. Sarmad Abdul Razak Abood Alsaadi, College of Medicine, Basic Sciences, Kirkuk University, Iraq
- 70) Dr. Abdulrahman Bello, Department of Veterinary Anatomy, Faculty of Veterinary Medicine, Usmanu Danfodiyo University, City Campus, Sokoto, Nigeria
- 71) Dr. Shahabaldin Rezania, Faculty of Civil Engineering (Environmental Engineering), Universiti Teknologi Malaysia (UTM), Malaysia

Publisher:

Excellent Publishers(Regd 71/2013) www.excellentpublishers.com email: excellentpublishers2013@gmail.com



Volume 5 Number 11 CONTENTS

November 2016

Original Research Articles

 Yasir Mohamed Abdelrahim, Hind Ahmed Ali, Mutaman Ali Abdelgader, Ahmed Adam Eisa, Elnur Elamin Abdelrahman and Eisa Ebrahim Elgaali
 Evaluation of Some Phytochemical Tests for Gumbail (Cordia africana Lam.) and its Uses in Termite Management
 Int.J.Curr.Microbiol.App.Sci.2016.5(11): 1-10
 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.001</u>
 [View Abstract] [View Full Text-PDF]

2. Arvind K. Sharma, Archana Srivastav and Asha Mukul Jana
Studies on Phytochemical Characteristics and Antimicrobial Activity of *Pleurotus spp*. Cultivated on Different Agro Wastes
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 11-23
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.002</u>
[View Abstract] [View Full Text-PDF]

3. R. Balasasirekha and P. Santhoshini Development of RTE Millet Mixes with Dehydrated Vegetable Peel Int.J.Curr.Microbiol.App.Sci.2016.5(11): 24-37 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.003</u> [View Abstract] [View Full Text-PDF]

4. Vijay Bharti and V. Santhalakshmi

A Comparative Study on Knowledge Regarding Labour Pain and Cultural Beliefs on Labour Pain Relief Measures among Rural and Urban Antenatal Women at Civil Hospitals Mohali, Punjab, India Int.J.Curr.Microbiol.App.Sci.2016.5(11): 38-44 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.004</u> [View Abstract] [View Full Text-PDF]

5. Manjot and V. Santhalakshmi

A Descriptive Study to Assess the Bio-Psychosocial Consequences of Late Marriages among Late Married Couples Residing in Selected Areas of District Mohali, Punjab, India Int.J.Curr.Microbiol.App.Sci.2016.5(11): 45-50 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.005</u> [View Abstract] [View Full Text-PDF]

6. Bhavini N. Rathod, Harshil H. Bhatt and Vivek N. Upasani
Extracellular Hydrolases producing Haloarchaea from Marine Salterns at Okhamadhi, Gujarat, India
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 51-64
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.006</u>
[View Abstract] [View Full Text-PDF]

7. Garima Bhardwaj and Bhagat Singh

Potential MIC of Bioactive Peptides from Fermented Bovine Milk to Inhibit Bacterial Pathogens

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 65-73 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.007</u> [View Abstract] [View Full Text-PDF]

8. Mehal Passi, Nakul Aggarwal and Anu Priya Minhas

Detecting *Cronobacter* Contamination in Protein Mixture and Biscuit Sample by Conventional PCR Method-A Preliminary Study Int.J.Curr.Microbiol.App.Sci.2016.5(11): 74-85 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.008</u> [View Abstract] [View Full Text-PDF]

9. M. Saleem, R. Gopal, T. Mangaiyarkarsi, S. Sunil, J. Krishnapriya and R. Nagma Profile of Bacterial Isolates from Blood Cultures and their Antimicrobial Susceptibility Pattern

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 86-91 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.009</u> [View Abstract] [View Full Text-PDF]

10. Hitesh R. Ahir and Hetal G. Vaghela

Seroprevalence of Dengue Viral Infection in Patients attending Tertiary Care Hospital, South Gujarat, India Int.J.Curr.Microbiol.App.Sci.2016.5(11): 92-26 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.010</u> [View Abstract] [View Full Text-PDF]

11. Bruna Polacchine da Silva, Josielle Abrahão and Rosane Marina Peralta

Effects of Carbon Sources and Time of Cultivation on the Antimicrobial Activities of Intra and Extracellular Extracts of *Pleurotus pulmonarius* Cultured in Submerged Conditions Int.J.Curr.Microbiol.App.Sci.2016.5(11): 97-105 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.011</u>

[View Abstract] [View Full Text-PDF]

12. Moustafa A. El-Nakeeb, Hoda M. Omar, Rania Abozahra and Eman Badr

Comparative Study of Different Methods of Detection of -lactamase Production in Gram Negative Bacterial Isolates and Studying Induction of Resistance to Cefipime Int.J.Curr.Microbiol.App.Sci.2016.5(11): 106-120 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.012</u>

[View Abstract] [View Full Text-PDF]

13. Pranaba Nanda Bhattacharyya, Prasanta Dutta, Satya Ranjan Sarmah, Amarjyoti Tanti, Mausomi Madhab and Rofika Begum

Pranaba Nanda Bhattacharyya, Prasanta Dutta, Satya Ranjan Sarmah, Amarjyoti Tanti, Mausomi Madhab and Rofika Begum

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 121-125

DOI: http://dx.doi.org/10.20546/ijcmas.2016.511.013

[View Abstract] [View Full Text-PDF]

14. Kavita Bandivadekar, Shruti Garge, Aditi Phand and Manjusha Bhave

Use of Plant Auxins Produced by Bacteria in Plant Tissue Culture and Seed Pre-treatment; A Possibility of Replacement of Synthetic Auxins Int.J.Curr.Microbiol.App.Sci.2016.5(11): 126-131

DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.014</u> [View Abstract] [View Full Text-PDF]

15. Rachana B. Chavan and Meenakshi B. Bhattacharjee
Role of Alginate and Oxalic acid in Ameliorating Se Toxicity in *Hapalosiphon* cyanobacterium
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 132-139
DOI: http://dx.doi.org/10.20546/ijcmas.2016.511.015
[View Abstract] [View Full Text-PDF]

16. Kirti Nirmal, Rumpa Saha, V.G. Ramachandran, Shukla Das and S.N. Bhattacharya Resistance in *Neisseria gonorrhoeae*: A Cause of Concern? Int.J.Curr.Microbiol.App.Sci.2016.5(11): 140-144 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.016</u> [View Abstract] [View Full Text-PDF]

17. Abhishek Mehta1, Gaurav Saxena and Abin Mani

Comparative Analysis of Antibacterial Activity of Aqueous, Ethanolic, Methanolic and Acetone Extracts of Commercial Green Tea and Black Tea against Standard Bacterial Strains Int.J.Curr.Microbiol.App.Sci.2016.5(11): 145-152 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.017</u> [View Abstract] [View Full Text-PDF]

18. Satisfaction Syntem, Himadri Dutta and M. Kalyani Characterization of *Proteus* Species and Detection of Multi Drug Resistant (MDR) with Special Reference to ESBL Strains Int.J.Curr.Microbiol.App.Sci.2016.5(11): 159-160 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.018</u> [View Abstract] [View Full Text-PDF]

19. Aya H. Alhalaby and Ashwak B. J. Al-Hashmiy
Assessment of Biofilm Production and Antibiotic Pattern in E. faecium and E. faecalis isolated from Some UTI Iraqi Patients
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 161-172
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.019</u>
[View Abstract] [View Full Text-PDF]

20. Vilasini Udyavara, Sangeetha Madambithara Sivadasan, Dechamma Mundanda Muthappa, Santhosh Kogaluru Shivakumaraswamy and Moleyur Nagarajappa Venugopal Prevalence and Genomic Characterization of *Vibrio parahaemolyticus* isolated from Molluscan Shellfish and their Inhabiting Water of Coastal Karnataka, India Int.J.Curr.Microbiol.App.Sci.2016.5(11): 173-182

DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.020</u> [View Abstract] [View Full Text-PDF]

21. Marwa M.E. Abd-Elmonsef, Naglaa F. Ghoname, Abeer Shahba, Maha M. Hagras, Lamees M. Dawood and Mohamed M.E. Abd-Elmonsef

Association of the RETN–420C G Polymorphism with Rheumatoid Arthritis in an Egyptian Population

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 183-195 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.021</u> [View Abstract] [View Full Text-PDF]

22. Poonam Jayant Singh and Atul Kumar Tiwari

The Ongole Cattle Germplasm: Corollary Arising out of Regulating Access, Associated Knowledge and Benefit Sharing Post Nagoya Protocol by Brazil from India Int.J.Curr.Microbiol.App.Sci.2016.5(11): 196-204 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.022</u> [View Abstract] [View Full Text-PDF]

23. J.Nasreen Banu and V. Gayathri

PreparationofAntibacterialHerbalMouthwashagainstOralPathogensInt.J.Curr.Microbiol.App.Sci.2016.5(11):205-221DOI:http://dx.doi.org/10.20546/ijcmas.2016.511.023[View Abstract][View Full Text-PDF]

24. Suzan F. Ali, Walaa Othman Elshabrawy, Muneera A. Alsheeha, and Ahmed Hussein Validity of Different Laboratory Methods for Diagnosis of Infectious Vaginitis: Real-Time Polymerase Chain Reaction for Detection of Trichomoniasis Int.J.Curr.Microbiol.App.Sci.2016.5(11): 222-229 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.024</u> [View Abstract] [View Full Text-PDF]

25. Hala B. Othman and Dalia H. Abd ElHamid

Evaluation of Phenotypic Methods for Detection of Plasmid-Mediated AmpC -Lactamases (PMABLs) among Klebsiella pneumoniae Int.J.Curr.Microbiol.App.Sci.2016.5(11): 230-239 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.025</u> [View Abstract] [View Full Text-PDF]

26. Abd-El Aziem Farouk, N. Thoufeek Ahamed, Othman AlZahrani, Khalid H. Alamer and Abdul Aziz Bahobail
Antimicrobial Activities Evaluation from the Extracts of Leaves, Flowers, Fruits and Latex of *Calotropis procera* from Taif
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 240-256
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.026</u>
[View Abstract] [View Full Text-PDF]

27. S.Saravanan and R. Sivakami

A Seasonal Analysis of Micro Zooplankton in the Backwaters of Kottaipattinam, Tamil Nadu, India

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 257-262 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.027</u> [View Abstract] [View Full Text-PDF]

28. Nadakuduru Premanadham, K. Jitendra, Munilaxmi, Siva Prasad Reddi, Ramya and Charan Kumar

Antibiotic Resistance Pattern of *Pseudomonas aeruginosa* Strains Isolated from Blood Cultures-Batec/Alert3D in a Tertiary Care Centre Narayana Hospital and Medical College Nellore AP, India

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 263-268 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.028</u> [View Abstract] [View Full Text-PDF]

29. P.R. Lyra, Sindhu Cugati, K. Anuradha and D. Venkatesha Metallo Beta Lactamase Producing *Pseudomonas aeruginosa* in a Tertiary Care Hospital Int.J.Curr.Microbiol.App.Sci.2016.5(11): 269-274 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.029</u> [View Abstract] [View Full Text-PDF]

30. Sarita Kumari, Anukool Vaishnav, Shekhar Jain, Devendra Kumar Choudhary and Kanti Prakash Sharma

Regulation of Ethylene Level in Mungbean (*Vigna radiata L.*) by 1-Aminocyclopropane-1-Carboxylic Acid (ACC)-Deaminase containing Bacterial Strain under Salt Stress Int.J.Curr.Microbiol.App.Sci.2016.5(11): 275-283 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.030</u> [View Abstract] [View Full Text-PDF]

31. Abd-ElAziem Farouk, N. Thoufeek Ahamed, Othman AlZahrani, Khalid H. Alamer, Yassin Al-Sodany and Abdul Aziz Bahobail
Antimicrobial Activity of *Caralluma quadrangula* (Forssk) N.E. Br Latex from Al-Shafa Taif, Kingdom of Saudi Arabia
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 284-298
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.031</u>
[View Abstract] [View Full Text-PDF]

32. Kalaivani Ramakrishnan, Arunava Kali, M.V. Pravin Charles and S. Seetha Kunigal Comparative Evaluation of Three Stool Concentration Techniques in the Diagnosis of Intestinal Parasitic Infections Int.J.Curr.Microbiol.App.Sci.2016.5(11): 299-304 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.032</u> [View Abstract] [View Full Text-PDF]

33. Rohan Gavankar and Maya Chemburkar Isolation and Characterization of Native Yeast from *Mahua* Flowers Int.J.Curr.Microbiol.App.Sci.2016.5(11): 305-314 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.033</u> [View Abstract] [View Full Text-PDF]

34. S.Singh, S.A. Samant, M. Bansal, A. Talukdar and D. Arif
Phenotypic Detection of Carbapenemase Producing Gram Negative Bacteria by Modified
Hodge Test
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 315-320
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.034</u>
[View Abstract] [View Full Text-PDF]

35. Prakash Kariyajjanavar, Sandesh Yargol, K.G. Somashekar Achar and T.R. Parashurama Traditional Healthcare Knowledge of Sedum Taluk, Kalaburgi, Karnataka, South India Int.J.Curr.Microbiol.App.Sci.2016.5(11): 321-328 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.035</u> [View Abstract] [View Full Text-PDF]

36. T.V. Parimala

A Retrospective Study on Seroprevalence of Hepatitis B Surface Antigen among Patients in a Tertiary Care Hospital, South India Int.J.Curr.Microbiol.App.Sci.2016.5(11): 329-333 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.036</u> [View Abstract] [View Full Text-PDF]

37. Dipali Gupta, Vijender Singh and Nipun Agrawal

Volatile Constituents and Antimicrobial Activities of Dried Rhizome of Cyperus otundus Linn.

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 334-339 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.037</u> [View Abstract] [View Full Text-PDF]

38. S.Umadevi, S. Pramodhini, Srirangaraj and K.S. Seetha
Resistance Pattern of *Enterococcus spp.* Isolated from Clinical Specimens
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 340-345
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.038</u>
[View Abstract] [View Full Text-PDF]

39. Indrajit Gupta, Purbasha Ghosh, Ayan Nandi, Prosenjit Naskar, Sayantan Mondal and G.D. Mitra

A Study on Identification and Antifungal Susceptibility Pattern of different Candida Species Isolated from Various Clinical Specimens in a Tertiary care Hospital of West Bengal, India Int.J.Curr.Microbiol.App.Sci.2016.5(11): 346-350 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.039</u> [View Abstract] [View Full Text-PDF]

40. S.K. Mengane Phytochemical Analysis of Adiantum lunulatum Int.J.Curr.Microbiol.App.Sci.2016.5(11): 351-356 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.040</u> [View Abstract] [View Full Text-PDF]

41. Mahesh Chitrakoti and Kalpana Chandramore
Phylogenetic Diversity of Bacterial Communities of Western India Oil Fields by PCR-DGGE
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 357-370
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.041</u>
[View Abstract] [View Full Text-PDF]

42. U.K. Ezemagu, C.I.P Anibeze, C.O. Ani and G.C Ossi

Correlation of Body Mass Index with Low Back Pain amongst Patients without Injury in a Nigeria Population

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 371-378 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.042</u> [View Abstract] [View Full Text-PDF]

43. Pivot S.A. Sachi, Innocent Yaou Bokossa, Célestin C.K. Tchekessi, Roseline M. Bleoussi, Jultesse S.B. Banon, Anayce A. Djogbe, Karl Assogba and Guy A. Mensah Assessment of the Bacteriological Quality of an Aquatic Ecosystem in South of Benin: Case of Ahémé Lake

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 379-387 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.043</u> [View Abstract] [View Full Text-PDF]

44. Anandhi Lakshmanan, Dharmalingam Thirunavukkarasu, T.Umaarasu and S. Rajesh
Assessment of Amp C Beta-lactamase Production by Various Methods in Urinary Isolates of *Escherichia coli* from a Tertiary Care Teaching Hospital
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 388-395
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.044</u>
[View Abstract] [View Full Text-PDF]

45. Baqur A. Sultan, Kafil Akhtar, Eman Jabbar Khadum, Safa-Aldeen S. Nema, Falah Dali and Rana K. Sherwani
Pathological and Therapeutic Association between Enterobiasis and Certain Perianal Problems
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 396-400
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.045</u>
[View Abstract] [View Full Text-PDF]

46. Adnan H. Muhammed Ali and Muhammed Ridha H. Shubber Modes of Nasopharyngeal Carcinoma Presentation Int.J.Curr.Microbiol.App.Sci.2016.5(11): 401-406 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.046</u>
[View Abstract] [View Full Text-PDF]

47. Bothaina Abd El Hakeem, Hanan A. Emarah and Sraa A. Abo Melha Relationship between Dietary Patterns and Body Mass Index of Pregnant Women in Khamis Mushayt, Saudi Arabia Int.J.Curr.Microbiol.App.Sci.2016.5(11): 407-415 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.047</u> [View Abstract] [View Full Text-PDF]

48. Pradnya Atmaram Jadhav and Shahriar B. Roushani
Prevalence of Vancomycin Resistant Enterococcus and its Antibiotic Resistance Pattern at a Tertiary Care Hospital
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 416-424
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.048</u>
[View Abstract] [View Full Text-PDF]

49. Shailaja Harugade, A.B. Gawate and Bharat Shinde
Bee Floral Diversity of Medicinal Plants in Vidya Pratishthan Campus, Baramati, Pune, District (M.S.) India
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 425-431
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.049</u>
[View Abstract] [View Full Text-PDF]

50. Eny Puspani, Desak Putu Mas Ari Candrawati and Dan I.G.N.G. Bidura Implementation Probiotics Cellulolitic B-7 Bacteria (Isolation from Buffalo Rumen) into Rations on the Performance, Abdominal Fat and Serum Cholesterol of Duck Int.J.Curr.Microbiol.App.Sci.2016.5(11): 432-441 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.050</u> [View Abstract] [View Full Text-PDF]

51. S. Prasanna, S. Dharanidevi, Nikunja Kumar Das and Sathya Raj
Prevalence, Phenotypic Characterization and Antibiotic Susceptibility of Non-Fermentative
Gram Negative Bacilli Isolates at a Tertiary Care Centre
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 442-454
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.051</u>
[View Abstract] [View Full Text-PDF]

52. D.A. Desai, G.P. Kukreja, C.J. Raorane and S. B. Patil
Partial Sequencing of Serendipitously isolated Antifungal Producer, *Pseudomonas tolaasii* Strain GD76 16s Ribosomal RNA Gene
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 455-458
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.052</u>
[View Abstract] [View Full Text-PDF]

53. Santanu Maitra

Study of Genetic Determinants of Nickel and Cadmium Resistance in Bacteria-A Review Int.J.Curr.Microbiol.App.Sci.2016.5(11): 459-471 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.053</u> [View Abstract] [View Full Text-PDF]

54. Ponnusamy Ponmurugan, Fahad Khalid Aldhafiri and Santhanaraj Balakrishnan Antibacterial Activity of Green Tea Leaves Int.J.Curr.Microbiol.App.Sci.2016.5(11): 472-477 DOI: http://dx.doi.org/10.20546/ijcmas.2016.511.054 [View Abstract] [View Full Text-PDF]

55. Jitendra Malviya and Vaibhavi Joshi

Anticancer Activity Evaluation and Some Indian Medicinal Plants from Amarkantak Mekal Plateau Madhya Pradesh, India Int.J.Curr.Microbiol.App.Sci.2016.5(11): 478-483 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.055</u> [View Abstract] [View Full Text-PDF]

56. D.Saikumari, S.K. Shiva Rani and Neeti Saxena Antibacterial Activity of *Syzigium aromaticum L*. (Clove) Int.J.Curr.Microbiol.App.Sci.2016.5(11): 484-489 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.056</u> [View Abstract] [View Full Text-PDF]

57. S.A. Patel and S.C. Parikh

Isolation and Screening of Probiotic Potential Lactic Acid Bacteria from Local Dairy Products Int.J.Curr.Microbiol.App.Sci.2016.5(11): 490-498 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.057</u> [View Abstract] [View Full Text-PDF]

58. Maasumeh Kaviani, Tahere Abdullahi Tehrani, Sara Azima, Khadijeh Abdali and Nasrin Asadi

Comparison of the Effects of Simultaneous Use of Methylergonovine and Combined Lowdose (LD) Contraceptive Pills on Hemorrhage Due to Retained Pregnancy Products after Abortion

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 499-507 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.058</u> [View Abstract] [View Full Text-PDF]

59. Vrutika V. Gandhi, P. Neelu Sree and M. Kalyani

A Study on Etiological Agents with Special Reference to Fungal Isolates causing Chronic Suppurative Otitis Media in a Tertiary Care Hospital Int.J.Curr.Microbiol.App.Sci.2016.5(11): 508-514 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.059</u> [View Abstract] [View Full Text-PDF]

60. Harekrishna Nath and Dipa Barkataki

Prevalence of ESBL and MBL producing Acinetobacter Isolates in Clinical Specimens in Tertiary Care Hospital, Assam, India Int.J.Curr.Microbiol.App.Sci.2016.5(11): 515-522 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.060</u> [View Abstract] [View Full Text-PDF]

61. H.S. Shilpa and J. Mariraj

Intestinal Parasitic Infections in Relation to HIV/AIDS Status, Diarrhoea and CD4 T-Cell Count

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 523-531

DOI: http://dx.doi.org/10.20546/ijcmas.2016.511.061 [View Abstract] [View Full Text-PDF]

62. Alex. M. Varghese Awareness and Attitude of Medical Negligence and Medical Ethics among Interns and

Resident Doctors Int.J.Curr.Microbiol.App.Sci.2016.5(11): 532-535 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.062</u> [View Abstract] [View Full Text-PDF]

63. Siva Prasad Reddy Basava, Srijan Ambati, Kandati Jithendra, N. Premanadham, P.Sreenivasulu Reddy and Charan Kumar Mannepuli Efficacy of Iodine-Glycerol versus Lactophenol Cotton Blue for Identification of Fungal Elements in the Clinical Laboratory

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 536-541 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.063</u> [View Abstract] [View Full Text-PDF]

64. Smita Kulshreshtha, Usha Verma, P.K. Khatri and Prameshwar Lal

Prevalence of Oral Candida Carriage Rate among HIV Infected Asymptomatic and Non Infected Persons, their Antimycotic Sensitivity and its Association with CD4 Counts Int.J.Curr.Microbiol.App.Sci.2016.5(11): 542-545 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.064</u> [View Abstract] [View Full Text-PDF]

65. Firyal N. Aziz

Evaluation of Antibiotics Sensitivities against Some UTI Pathogens Int.J.Curr.Microbiol.App.Sci.2016.5(11): 546-551 DOI: http://dx.doi.org/10.20546/ijcmas.2016.511.065 [View Abstract] [View Full Text-PDF] 66. S.P. Adhikary and S.N. Sabat Impact of Cocklebur (*Xanthium indicum*) Allelochemicals on Vegetative Growth Parameters of Green Gram (*Phaseolus radiatus L.*) Int.J.Curr.Microbiol.App.Sci.2016.5(11): 552-564 DOI: http://dx.doi.org/10.20546/ijcmas.2016.511.066 [View Abstract] [View Full Text-PDF]

67. Madhuri R. Basutkar, Channappa T. Shivannavar and Subhaschandra M. Gaddad Biodegradation of Reactive Red-11 by the Isolate *Enterococcus casseliflavus* CMGS-1 Strain Int.J.Curr.Microbiol.App.Sci.2016.5(11): 565-578 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.067</u> [View Abstract] [View Full Text-PDF]

68. J.Radhakrishna Rao, V. Gireesh Kumar, T. Madhava Rao, D.Pramod Kumar, K. Chandra Shekar Reddy and K.B.P. Raghavender Use of Hybrid External Fixation Technique in the Repair of Long Bone Fractures in Dogs Int.J.Curr.Microbiol.App.Sci.2016.5(11): 579-586 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.068</u>

[View Abstract] [View Full Text-PDF]

69. Nabila Akbar and M. Kalyani
Clinico–Mycologicalal Profile of Dermatophyte Species in a Tertiary Care Center
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 587-593
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.069</u>
[View Abstract] [View Full Text-PDF]

70. J. Meenakshi and V. Kalai Gandhi

Development of Herbal-Silver Nano Composite (HSNC): Antibacterial Evaluation and Investigation of *fnb* A and *fnb* B Genes Coding Fibronectin-binding Proteins in *Staphylococcus aureus*

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 594-603 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.070</u> [View Abstract] [View Full Text-PDF]

71. Andrew A. Minamor and G.T. Odamtten

Radial Growth of three *Paecilomyces* Species Isolated from Two Ghanaian Maize Varieties Abeleehi and Obaatanpa on Five Different Media and the Effects of their Culture Filtrate on Seed Germination and Radicle Elongation of Abeleehi and Obaatanpa

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 604-617 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.071</u> [View Abstract] [View Full Text-PDF]

72. Samaneh Mousavi, Abbas Yazdanpanah and Irvan Massoudi Asl

Comparing Quality of Life (QoL) during Pregnancy among Pregnant Women Referring to Public or Private Hospitals

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 618-622 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.072</u> [View Abstract] [View Full Text-PDF]

73. Neelam K. Naranje, Bharat J. Wadher and Manohar G. Muddeshwar
Utilization of Low Cost Carbon Substrates and Optimization Studies for Maximum Yield of Polyhydroxybutyrate from B.megaterium
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 623-630
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.073</u>
[View Abstract] [View Full Text-PDF]

74. Rajdeep Paul, Shyam Sundar Bera and Sumi Nongrum A Review of Advance Techniques in Diagnosis of Malaria Int.J.Curr.Microbiol.App.Sci.2016.5(11): 631-636 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.074</u> [View Abstract] [View Full Text-PDF]

75. Rasha M. Abdel Hamid, Safaa S. Hassan, Hadir A. El-Mahallawy and Magdy Saber Molecular Characterization of Carbapenem Resistant *Acinetobacter baumannii* in Cancer Patients Int.J.Curr.Microbiol.App.Sci.2016.5(11): 637-647 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.075</u> [View Abstract] [View Full Text-PDF]

76. V. Mangayarkarasi, Kanishka Hrishi Das, Shabana Praveen, V.Chitraleka and Christopher Amalraj
Fluconazole Resistance Pattern of Candida isolates in Clinical Samples by E-Test
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 648-655
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.076</u>
[View Abstract] [View Full Text-PDF]

77. Bob I.A. Mgbeje, Terungwa A. Kalana, Flora O. Ugoanyanwu and Patrick E. Ebong
Effect of Phytochemical Fractions of Vernonia amygdalina on Liver of STZ Induced Diabetic
Male Wistar Rats
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 656-667
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.077</u>
[View Abstract] [View Full Text-PDF]

78. A.S. El-Hassanin, M.R. Samak, Moustafa, N. Shafika, A.M. Khalifa and M. Ibrahim Inas Effect of Foliar Application with Humic Acid Substances under Nitrogen Fertilization Levels on Quality and Yields of Sugar Beet Plant Int.J.Curr.Microbiol.App.Sci.2016.5(11): 668-680 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.078</u>
[View Abstract] [View Full Text-PDF]

79. Arun Kumar Sharma, Vinay Sharma and Jyoti Saxena
Characterization of Lipase from Wild (LPF-5) and Mutant (HN1) Strain of Aspergillus nigera
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 681-690
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.079</u>
[View Abstract] [View Full Text-PDF]

80. Mohamed A. EL-Hag, Eman F. Sharaf and Medhat A. El-Naggar Amplitude Modulated Waves as a Promising Tool for Food Preservation Int.J.Curr.Microbiol.App.Sci.2016.5(11): 691-702 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.080</u> [View Abstract] [View Full Text-PDF]

81. P. Kanchanadevi and S. Chandra Sekaran
Importance of EDTA in the Detection of Metallo Beta Lactamase from Imipenem Resistant
Gram Negative Bacilli
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 702-706
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.081</u>
[View Abstract] [View Full Text-PDF]

82. Vijaykumar Biradar
Extraction of Phytochemicals from Local Selected Plants and their Antibacterial Role
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 707-720
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.082</u>
[View Abstract] [View Full Text-PDF]

83. Sonali L. Parekh, Smitha Balakrishnan, Subrota Hati and K. D. Aparnathi Lactulose: Significance in Milk and Milk Products Int.J.Curr.Microbiol.App.Sci.2016.5(11): 721-732
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.083</u>
[View Abstract] [View Full Text-PDF]

84. Nitin Arun Ambhore, Poonam C. Sayare, Rajesh P. Karyakarte and Rupali S.Mantri Trends of Seroprevalence and Epidemiology of HIV Infection: A Five Year Tertiary Care Hospital Based Study
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 733-740
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.084</u>
[View Abstract] [View Full Text-PDF]

85. Varunika Vijayvergia, Sunita Gupta and Jaswant Goyal
Neonatal Septicemia - Bacteriological Spectrum and Antibiogram-A Study from a Tertiary
Care Center of North India
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 741-750
DOI: http://dx.doi.org/10.20546/ijcmas.2016.511.085
[View Abstract] [View Full Text-PDF]

86. Gatot Siswo Hutomo, Abdul Rahim and Syahraeni Kadir
Pectin Isolation from Dry Pod Husk Cocoa with Hydrochloride Acid
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 751-756
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.086</u>
[View Abstract] [View Full Text-PDF]

87. Azher Hameed Al-Taie, Abdulnabi Abbdul Ameer Matrood and Muhammed Al-asadyi The Influence of Some Fungi Bio-genic on Promoting Growth and Yield of Wheat-Var. Ibaa99

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 757-764 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.087</u> [View Abstract] [View Full Text-PDF]

88. Ranbir Singh, V.K. Razdan and Manika Sharma
Prevalence and Detection of Tomato Leaf Curl Virus from Low Altitude Subtropical Areas of Jammu and Kashmir
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 768-773
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.088</u>
[View Abstract] [View Full Text-PDF]

89. Hissah Al-Turki, Samirah Al-Ali, Reem Al-Khaldi, Muneerah Al-Rowaila, and Lorina Badger-Emeka

In Vivo Studies comparing the Fever Patterns using *Escherichia coli* and *Pseudomonas aeruginosa as*Pyrogens in Rabbits Int.J.Curr.Microbiol.App.Sci.2016.5(11): 774-781

DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.089</u> [View Abstract] [View Full Text-PDF] 90. Utsav, Baidyanath Kumar and Atul Kumar

Efficacy of *Circuma longa Linn*. Extract in the Management of Type-2 Diabetes Mellitus associated Hypercholesterolemia in Mice Diabetic Model Int.J.Curr.Microbiol.App.Sci.2016.5(11): 782-794 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.090</u> [View Abstract] [View Full Text-PDF]

91. Utsav, Baidyanath Kumar and Atul Kumar

Diabetes Mellitus and its Control by *Ocimum sanctum* Extract in Mice Diabetic Model Int.J.Curr.Microbiol.App.Sci.2016.5(11): 795-810 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.091</u> [View Abstract] [View Full Text-PDF]

92. A.A.A. Mekdad and A.M.A. El-Sherif

The Effect of Nitrogen and Potassium Fertilizers on Yield and Quality of Sweet Sorghum Varieties under Arid Regions Conditions

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 811-823

DOI: http://dx.doi.org/10.20546/ijcmas.2016.511.092

[View Abstract] [View Full Text-PDF]

93. Aziz Fatima, Farah Malik, Amna Shafiq, Sumara Jawaid, Shazia Tabassum Hakim and Sayyada Ghufrana Nadeem

Evaluation Antibacterial Activity of three Most Consumed Tea Extracts against Pathogenic Bacteria

Int.J.Curr.Microbiol.App.Sci.2016.5(11): 824-827 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.093</u> [View Abstract] [View Full Text-PDF]

94. Adeela Iqbal, M. Shafi Hasni, Sajjad-ur-Rahman, Rizwan Aslam and Kasib Khan Preparation and Evaluation of Bacteriophage Lysate Specific for *Salmonella typhimurium* Int.J.Curr.Microbiol.App.Sci.2016.5(11): 829-835 DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.094</u> [View Abstract] [View Full Text-PDF]

95. Hanan M. Abobaker, Hamida EL. Elsalhin and Hamad M. Adress Hasan Nitrogen, Phosphorus and Minerals (Sodium, Potassium and Calcium) Contents of Some Algae's Species (Anabaena and Spirulina platensis)
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 836-841
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.095</u>
[View Abstract] [View Full Text-PDF]

96. Ojo-Omoniyi, Olusola Abayomi, Okwa, Omolade Olayinka, Junaid, Iswat Olaide and Ikuoye, Abosede Oyebola
Sustainable Solid Waste Management: Isolation of Cellulolytic Microorganisms from Dumpsites in Lagos, Southwest Nigeria
Int.J.Curr.Microbiol.App.Sci.2016.5(11): 842-853
DOI: <u>http://dx.doi.org/10.20546/ijcmas.2016.511.096</u>
[View Abstract] [View Full Text-PDF]



International Journal of Current Microbiology and Applied Sciences ISSN: 2319-7706 Volume 5 Number 11 (2016) pp. xx-xx Journal homepage: <u>http://www.ijcmas.com</u>



Original Research Article

http://dx.doi.org/10.20546/ijcmas

Implementation Probiotics Cellulolitic B-7 Bacteria (Isolation from Buffalo Rumen) into Rations on the Performance, Abdominal Fat and Serum Cholesterol of Duck

Eny Puspani*, Desak Putu Mas Ari Candrawati and Dan I.G.N.G. Bidura

Faculty of Animal Science, Udayana University, Denpasar, Bali, Indonesia Jl. PB. Soedirman, Denpasar, Bali-Indonesia *Corresponding author email id: <u>eny_fapet@yahoo.co.id</u>

ABSTRACT

Keywords

Probiotics, buffalo rumen, performance, cholesterol, abdominal-fat, duck

Article Info

Accepted: xx October 2016 Available Online: xx November 2016 This research aims to study the influence of cellulolitic B-7 bacterial isolates rumen of buffalo as a source of probiotics on the performances, abdominal fat, and cholesterol serum of a male Bali duck. A total of 240 male bali ducks were randomized in a completely randomized design (CRD) with four treatments and six replications. Treatment is attempted is the ducks that were given rations without supplementation of probiotic cultures cellulolitic as control (A); ration with 0.20% (B); 0.40%; and 0.60% probiotic cultures selulolityc B-7, resvectively. The isolate has passed the test and has been considered as potential probiotic according to our previous study (Bidura et al., 2014). The results showed that the ducks were given rations containing probiotics result in weight gain and feed efficiency significantly (P<0.05) better than the control. However, administration of probiotics through diet significantly (P<0.05) can reduce abdominal fat weight and serum kolseterol in ducks. From these results it can be concluded that supplementation of 0.20% to 0.60% probiotic cellulolitic B-7 bacteria were isolated from the rumen of buffalo in the ration can significantly improve live weight gains and feed efficiency of Bali duck. In contrast, significantly reduced abdominal-fat weight and serum cholesterol of ducks.

Introduction

Public attention to the fat and cholesterol become greater, especially after it emerged that the consumption of fat/cholesterol excess will affect health. Not only to the increase in coronary heart disease, but lately informed also against cancer, diabetes, and high blood pressure (Santoso and Sartini. 2001). It was also reported that accumulation of high fat in the stomach and viscera may reduce the benefits of post-

utilizing probiotics. The use of antibiotics to stimulate the growth of poultry has been banned in European and American countries (Ahmad,

harvest plant and improve wastewater

treatment problem. Therefore, it is very

helpful if it can reduce cholesterol and

abdominal fat chicken with biotechnology

2006). Therefore, nutritionists and livestock

production attracted to compounds that can serve as a substitute for antibiotics. Probiotics is one approach that has the potential to replace antibiotics. Probiotics are live microorganisms which when administered through the digestive tract, have a positive impact on the health and production of the host. According to Putra *et al.* (2015), the addition of probiotics in the ration is expected to enhance the role of normal flora in the digestive tract of cattle to produce exogenous enzymes, such as amylase, protease, and lipase which can enhance the activity of endogenous enzymes to hydrolyze feed nutrients.

Probiotics contain beneficial bacteria species that is normally found in the intestinal tract. Studies show that consumption of certain probiotic strains, particularly Lactobacillus can alter the intestinal microflora to produce a beneficial effect. Most commercially available strains of *Lactobacilli* and *Bifidobacteria* species are generally considered safe and can be very helpful in of pediatric the treatment diarrhea. However, the clinical benefits of probiotic therapy depends on many factors, such as the type of bacteria, the accuracy of the dose, method of delivery, and other environmental factors underlying (Dinkci et al., 2006).

Based on this, interesting to note is the use of cellulolytic microbes from buffalo rumen fluid as the fiber degrading in the gastro intestinal tract of poultry. This is possible because the buffalo rumen fluid microbes turns out to have the highest cellulolytic activity compared with other livestock cellulolytic microbes. According to Sudirman (2004) in addition to determining the source of the microbial digestion of fiber activity, is also determined by the exact inoculum dose, type, microbial and microbial populations were used. Provision of microbial cultures from buffalo rumen fluid to the ducks is expected to lead to a synergistic effect between species of buffalo rumen microbes with microbes of duck in digestive tract, which can lead to the ability to digest fiber.

Some of the results of the preliminary study on the use of probiotics in the ration was able to improve performance, digestibility, efficiency ration, and lowering the amount of abdominal fat and cholesterol body poultry (Bidura et al., 2012; Bidura et al., 2014; Bidura et al., 2016; Candrawati et al., 2014). According to Siti et al. (2016), 0.40 supplementation of to 0.60% cellulolytic bacteria culture isolated from the rumen of buffalo in the tofu-based rations can improve the performance, carcass carcass percentage, weight, and the percentage of breast meat of ducks. In contrast, significantly reduced the amount of abdominal fat, blood serum cholesterol levels, and the concentration of N-NH3 in excreta of duck. Furthermore, the beneficial effect of supplementation of probiotics has also been reported by some research workers such as Yousefi and Karkoodi (2007), Yamada and Sgarbieri (2005), Queiroz et al. (2004) and Roni et al. (2014), Ghasemi et al. (2006), Mohiti et al. (2007). On the other hand, some authors such as Ayanwale et al. (2006)reported that probiotics supplementation on diet has no effect on performances.

Based on the description above research aimed to study the effect of supplementation cellulolytic bacteria selected from the rumen of buffalo into diets to improve performance and reduce levels of cholesterol of ducks.

Materials and Methods

Animals and experimental design

A total number of 240 male Bali duckling (*Anas sp*) at 2 weeks of age and with

homogenous body weight were randomly divided and caged in 24 separate pens with 10 birds in each pen. Prior to introduction of the experimental birds, the pens were thoroughly cleaned and disinfected. During the course of experiment, the birds were maintained under standard management condition for 42 days (6 weeks). For the purpose of examination of their carcasses and cholesterol concentration, 30 birds from each treatment group were randomly sampled.

A completely randomized design (CRD) was employed in the current experiment. Four treatment groups were assigned depending on their diets. All animals were fed an isonitrogenic and isocaloric ration in the form of mash and has been prepared in a such that meet the birds requirement to NRC (1984). Various levels of cellulolitic B-7 bacteria isolates obtained from rumen of buffalo as a source of probiotics slaughtered at the local abattoir, were supplemented on the diet. The difference in its level of application determined the four different experimental groups assigned. In the current experiment, addition of 0.0% (0 g/kg diet); 0.20% (2.0 g/kg diet); 0.40% (4.0 g/kg diet); and 0.60% (6.0 g/kg diet) were considered as the Treatment A (control), Treatment B, Treatment C, and Treatment D, respectively. Each group consisted of 6 replications with 10 birds in each replicate; therefore, the total number of experimental animals used in the current study was 240 birds. They were all provided with feed and drinking water ad libitum.

The probiotic used in the current study was cellulolitic B-7 bacteria isolates, which was isolated from from rumen of buffalo slaughtered at the local abattoir. The isolate has passed the test and has been considered as potential probiotic according to our previous study (Bidura *et al.*, 2014).

Body Composition

At the end of the experiment (56 days of age) 30 birds from each treatment were selected and slaughtered for determination of body composition. The leg and breast meats were separated from the carcass. The parts of the body fat are: pad-fat (separated from the organs of the abdominal viscera to the skin), mecenteric-fat (linkage separated from the intestine), vernticulus-fat, and abdominal-fat (a combination of fat-pad, ventriculus-fat, and mecenteric-fat).

Total serum cholesterol

For analysis of total serum cholesterol, two ml of blood was taken from the jugular vein of each duckling and centrifuged at 3000 rpm for 20 minutes. The blood samples were allowed to clot in a sample bottle and serum harvested was used to determine the total serum cholesterol content with the aid of a analyzed by the method of Liberman-Burchard (1980).

Statistical analysis

Data collected was subjected to Analysis of Variance and if significant different among the treatment group was noted, they were then underwent further statistical analysis following Duncan's Multiple Range Test (Steel and Torrie, 1989).

Results and Discussion

Results of the current study are presented in Table 1. It can be noted that supplementation on diet of various level of the probiotic ranging from 0.20% to 0.60% resulted in a significant (P<0.05) increase in live weight gains. Moreover, the supplementation of 0.20-0.60% of *cellulolytic B-7* bacteria culture significantly (P<0.05) increased feed efficiencies.

However, no significant different (P>0.05) in feed consumption was noted.

At the end of the experiment (at eight weeks of age), showed live weight gains during the six weeks of observation ducks that were given rations without supplementation of probiotic cellulolytic B-7 bacteris isolated from rumen buffalo as a control is 1058.75 g/birds (Table 1). Supplementation cellulolytic B-7 bacteria at the levels: 0:20%; 0:40%, and 0.60% in the ration, respectively were 18.48%; 19.56%; and 22.91% higher than controls and were statistically significantly different (P<0.05).

The mean value of the feed conversion ratio (feed consumption: weight gain) on ducks control is 5.45 (Table 1). The mean value of FCR ducks received treatment B, C, and D, respectively, were: 14.13%; 14.50%; and 16.88% (P<0.05) lower than the control. The results showed that the average amount of abdominal fat in ducks control was 2.85% body weight. The average amount of abdominal fat ducks received treatment B, C, and D respectively, are: 17.54%; 16.14%; and 18.60% (P<0.05) lower than the control. Total cholesterol levels in the blood serum of control duck was 185.64 mg dl (Table 1). Mean serum cholesterol levels of ducks treatment B, C, and D, respectively, were: 11.22%; 10.61%; and 12.70% (P<0.05) lower than the control.

The results showed that supplementation of 0.40 to 0.60% bacterial culture isolates cellulolitic B-7 were isolated from the rumen of buffalo in the diet can increase the performance of ducks (live weight gains and feed efficiency). Because culture isolate cellulolytic rumen of buffalo that have passed the test as an agent of probiotics and have activity cellulolitic or has an activity of CMC-ase (Bidura *et al.*, 2014) in the digestive tract is able to improve the

digestibility of the feed, in particular feed fiber roughness height, into compounds simpler so more easily absorbed by the body ducks.

Feed efficiencies and LWGs in bird offered probiotic group are higher than control groups. This may be caused by the fact that probiotics contain species of beneficial bacteria that are commonly found in the intestinal tract. Most commercially available strains of Lactobacilli and Bifidobacteria species are generally considered safe and may be especially helpful in treatment of diarrheal illnesses. However, clinical benefit of probiotic therapy is dependent on numerous factors such as type of bacteria, dosing regimen, delivery method, and other underlying host factors (Dinkçi et al., 2006). The use of cultures such as Saccharomyces cerevisiae can improve weight gain, as a result of the response to in creased dry matter intake. Especially, Saccharomyces cerevisiae, have been used in animal diets for several decades and are considered sources high quality proteins and B-complex vitamins, selenium and zince (Queiroz et al., 2004).

Moreover, Piao et al. (1999) noted that the use of 0.10% yeast (Saccharomyces serevisiae) in the ration significantly increased weight gain, feed efficiency and nutrient digestibility as well as reduce the amount of N and P secreted in the feces. Study in ducks by Bidura et al. (2012) also confirmed previous findings; the supplementation of 0.10% yeast culture led to an improvement in feed intake, feed conversion ratio, and weight gain. Such a finding is further supported by results of Umiarti et al. (2014) who found that the use of 0.10% to 0.30% Saccharomyces spp culture significantly improved live weight gains and feed efficiency. More positive effects of yeast supplementation were

reported by Mulyono *et al.* (2009), Wu *et al.* (2005) and Huang *et al.* (2004). Santin *et al.* (2001) found that supplementation of feed with *S.cerevisiae* cell wall (0.2%) improved broiler body weight. Many studies indicate that the addition of the culture of probiotic or enzyme in high feed content NSP can significantly reduce the viscosity of the digestive tract (intestinal viscosity), increase energy and protein retention (Wang *et al.*, 2004; Bidura *et al.*, 2012; Chen *et al.*, 2005).

According to Stanley et al. (1993), which found that broiler chickens were given probiotics 0.10% can significantly improve weight gain and feed efficiency. Giving feed containing probiotics can stimulate the metabolism of poultry feed in the digestive (Nurhayati, Probiotic process 2008). microbes such as Aspergillus oryzae and Saccharomyces cerevisiae in the digestive tract of poultry at the level of 0.15% and 0.30% can increase the activity of amylolytic and proteolytic enzymes, thus increasing energy and digestibility of dietary protein termetabolis (Han et al., 1999).

Apparently, the role of culture isolate cellulolytic rumen of buffalo as a probiotic agent in the digestive tract of ducks effective at levels from 0.20 to 0.60% in the ration. Piao et al. (1999) reported that probiotic supplementation in the diet can markedly increase weight gains. live nutrient absorption, and the absorption of nitrogen and phosphorus. The results of this study are supported by Wu et al. (2005) and Huang et (2004)that Aspergillus xylanase al. supplementation in wheat bran-based ration can improve the performance of broiler chickens. The same thing was reported by Mulyono et al. (2009) that the addition of 1.0% S.cerevisiae (9 x 10^9 cfu) derived from baker's yeast in the basal ration of broiler chickens significantly increase the digestibility of dry matter, digestible protein,

and protein efficiency ratio. Yi et al. (1996) reported that microbes in feed supplementation memperbaiiki can Ν retention in broiler and improve the digestibility of protein. It was also reported by Chen et al. (2005), that the addition of probiotic complex (L.acidophilus and S.cerivisae) in the basal diet can markedly increase the digestibility of dry matter feed.

Utama (2011),reported that the administration of probiotics (yeast Saccharomyces cerevisiae) in feed can improve the digestibility of protein and crude fiber components, such as cellulose and hemicellulose, since been overhauled in the form of simple monosaccharide. Chen et al. (2005) reported that the complex probiotic supplementation in the ration can markedly increase the digestibility of nutrients. Many studies indicate that the addition of culture probiotic or enzyme in high feed content NSP's can significantly reduce the viscosity of the digestive tract (intestinal viscosity), increase energy and protein retention (Wang et al., 2004; Bidura et al., 2012; Yi et al., 1996; Chen et al., 2005).

Supplementation of 0.20 to 0.60% bacterial culture isolates cellulolitic B-7 were isolated from the rumen of buffalo in the diet significantly reduced the amount of abdominal fat and blood cholesterol levels of ducks. These results are supported by Nurhayati (2008) that the use of the feed mixture fermented by Aspergillus niger at the level of 10-30% significantly reduced abdominal fat weight. Lactic acid bacteria in the digestive tract of poultry is able to utilize the energy derived from carbohydrates to lower the pH of the digestive tract to 4.5 resulting in the digestive tract becomes acidic. Acidic conditions causing lipase activity to be limited, so that the reduced fat digestion and

subsequent synthesis of fat to decrease. Research on pigs conducted by Min (2006) reported that feeding fermented can reduce the fat content.

Supplementation of culture isolates B-7 cellulolytic bacteria isolated from the rumen of buffalo in the ration significantly reduced plasma cholesterol levels. According Ezema and Yeh (2015), probiotics may contribute to the regulation of serum cholesterol concentrations were carried out by deconjugated bile acids. Cholesterol is a precursor to the formation of bile acids and

bile acid excretion when deconjugated enhanced by probiotic supplementation, the precursor molecule then be required to restore the formation of bile acids. The results of this study are supported by Park *et al.* (2008) and Sutarpa *et al.* (2011) reported that the use of probiotics in the diet can lower cholesterol levels in serum and chicken meat. Probiotic supplementation at a rate of 1.0 g/kg ration can be recommended for optimum egg production and to reduce the concentration of cholesterol in serum and egg (Ezema and Eze, 2015).

Table.1 Implementation of Probiotics cellulolitic B-7 bacteria isolates of Rumen Buffalo in rations for Reducing abdominal-fat and Cholesterol serum of Ducks (aged 2-8 weeks)

Variable	Supplemented of cellulolytic B-7 bacteria in rations (%)				SEM ¹⁾
	0.00	0.20	0.40	0.60	
Live weight gains (g/6 weeks)	1058.75b	1254.42a	1265.92a	1301.32a	52.056
Feed consumption (g/6 weeks)	5765.35a	5869.83a	5904.61a	5895.74a	78.457
Feed conversion ratio (feed/gains)	5.45a	4.68b	4.66b	4.53b	0.214
Abdominal fat (% berat badan)	2.85a	2,35b	2.39b	2.32b	0.107
Cholesterol serum (mg/dl)	185.64a	164.82b	165.95b	162.06b	5.038

Note:

1. Standart Error of the treatments means

2. Means with different superscripts within raws are significantly different (P<0.05)

Probiotic may contribute in the regulation of serum cholesterol concentrations conducted by deconjugated bile acids. As cholesterol is a precursor for bile acid formation and when deconjugated bile acids excretion is enhanced by probiotics supplementation, then more precursor molecules are needed for the recovery of bile acid formation (Ezema and Eze, 2015). Consequently, it may be expected that level of serum cholesterol decreases (Park *et al.*, 2008; Sutarpa *et al.*, 2011). Moreover, Klaver and Van Der Meer (1993) also suggested that co-precipitation with bile acids may be of importance in decreasing serum cholesterol concentrations. Ezema and Eze (2015) suggested, probiotic (*S.cerevisiae*) inclusion level of 1.0 g/kg of layers mash is rekomanded for optimum hen-day egg performance and minimum serum and egg cholesterol content (Bidura *et al.*, 2016).

DeSmet *et al.* (1998) reported that probiotics could contribute to the regulation of serum cholesterol concentration by deconjugated bile acids. Since the excretion of deconjugated bile acid is enhanced and cholesterol is it's precursor, more molecules are spent for recovery of bile acids (Ezema and Eze, 2015). As a result of increased synthesis of these acids, it is expected that the level of serum cholesterol will be reduced. That co-precipitation with bile acid might be of importance in decreasing serum cholesterol concentrattion.

In conclution, that supplementation of 0.20 to 0.60% cellulolytic B-7 bacteria culture isolated from the rumen of buffalo in rations can improve the performance of ducks. In contrast, significantly reduced the amount of abdominal fat and blood serum cholesterol levels of duck.

Acknowledgements

The authors would like to thank to staff of laboratory attendants at the Nutrition Laboratory, Udayana University for their assistance in chemical analysis of the samples. We also would like to deliver our appreciation to the Head of Research and Public Service Department and Rector of Udayana University, Denpasar for their support during the course of competition to obtain research fund.

References

- Ahmad, I., 2006. Effect of probiotics on broilers performance. Int. Poult. Sci., 5: 593-597.
- Ayanwale, B.A., M. Kpe and V.A. Ayanwale. 2006. The effect of supplementing *Saccharomyces cerevisiae* in the diets on egg laying and egg quality characteristics of pullets. Int. Poult. Sci., 5: 759-763.
- Bidura, I.G.N.G., I.B.G. Partama, D.K.H. Putra and U. Santoso. 2016. Implementation on diet of Probiotic Saccharomyces spp.SB-6 isolated from colon of Bali cattle on egg production and egg cholesterol

concentration of Lohmann brown laying hens. Int. J. Curr. Microbiol. App. Sci. 5 (4): 793-802. doi: http://dx.doi.org/

10.20546/ijcmas.2016.504.091

- Bidura, IGNG., I.B. Sudana, I.P. Suyadnya,
 I.G. Mahardika, I.G.L.Oka, I.B.Gaga
 Partama, and I.G.A.I. Aryani. 2012.
 The implementation of *Saccharomyces* spp.n-2 isolate culture (isolation from traditional yeast culture) for improving feed quality and performance of male
 Bali duckling. Agricultural Science
 Research Journal Vol. 2 (9): 486-492
- Bidura, IGNG., N.W. Siti and I. A. Putri Utami. 2014. Isolation of cellulolytic bacteria from rumen liquid of buffalo both as a probiotics properties and has CMC-ase activity to improve nutrient quality of soybean distillery byproduct as feed. International Journal of Pure & Applied Bioscience September Vol. 2 (5): 10-18
- Candrawati, DPMA., DA. Warmadewi and IGNG. Bidura. 2014. Implementation of *Saccharomyces spp.S-7* isolate (Isolated from manure of Bali cattle) as a probiotics agent in diets on performance, blood serum cholesterol, and ammonia-N concentration of broiler excreta. International Journal of Research Studies in Biosciences Vol. 2 (8): 6-16
- Chen, Y. J., K. S. Son, B. J. Min, J. H. Cho, O. S. Kwon, and I. H. Kim. 2005.
 Effects of Dietary Probiotic on Growth Performance, Nutrients Digestibility, Blood Characteristics and Fecal Noxious Gas Content in Growing Pigs. Asian-Aust. J. Anim. Sci. Vol. 18 (10): 1464-1468
- De Smet, I., L. Van Hoorde De Saaeyer, M. Van de Woeslyne and W. Verstraele. 1998. Cholesterol lowering in pig through enhanced bile salt hydrolase' activity. Br. J. Nutr., pp: 185-194

- Dinkçi, N., G. Ünal, S. Akalin and S. Gönç.2006. The Importance of Probiotics in Pediatrics. Pakistan Journal of Nutrition 5 (6): 608-611
- Ezema, C. And D.C. Eze. 2015. Probiotic effect (Saccharomyces of yeast cerevisiae) hen-day on egg performance, serum and egg cholesterol in laving chickens. Pakistan Journal of Nutrition 14 (1): 44-46
- Ghasemi, H.A., A.M. Tahmasbi, G.H. Moghaddam, M.Mehri, S. Alijani E. Kashefi and A. Fasifi, 2006. The effect of phytase and *Saccharomyces cerevisiae* (SC47) supplementation on performance serum parameters, phosphorous and calcium retention of broiler chickens. Int. Poult. Sci., 5: 162-168.
- Han, S.W., K. W. Lee, B. D. Lee and C. G.
 Sung 1999. Effect of Feeding Aspergillus oryzae Culture on Fecal Microflora, Egg Qualities, and Nutrient Metabolizabilities in Laying Hens. AJAS 12 (3): 417-421
- Huang, M. K., Y. J. Choi, R. Houde, J. W.
 Lee, B. Lee, and X. Zhao. 2004. Effect of *Lactobacilli* and *Acidophilic*Fungus On The Production Performance and Immune Responses
 In Broiler Chickens. Poult. Sci. 88: 788-795
- Lieberman, A. and R. Burchard, 1980. Enzimatic method to determined cholesterol. Engl. J. Med., 271: 915-924.
- Min, B. J. 2006. "Nutritional Value of Fermented Soy Protein (FSP) and Effect of FSP on Performance and Meat Quality of Pigs". (Ph.D. Thesis). Seoul, Korea: Department of Animal Resourches and Science.
- Mohiti, Asli, M., S.A. Hosseini, H. Ltfollahian and F. Shariatmadari, 2007. Effect of probiotics, yeast,

vitamin C supplements on performance and immune response of laying hen during high environmental temperature. Int. J. Poul. Sci., 6: 895-900.

- Mulyono, R. Murwani, dan F. Wahyono. 2009. Study Use of probiotic *Saccharomyces cerevisiae* as an Alternative to Antibiotics Additives to improve the usefulness of Protein and Energy in Broiler Chickens. J. Indonesian Trop. Anim. Agric. 34 (2): 145-151
- National Research Council. 1984. Nutrient Requirement Of Poultry. Washington, D. C.: National Academy Press.
- Nurhayati 2008. Pengaruh Tingkat Penggunaan Campuran Bungkil Inti Sawit Dan Onggok yang Difermentasi dengan Aspergillus Niger dalam Pakan terhadap Bobot dan Bagian-Bagian Karkas Broiler. Animal Production Vol 10 (1): 55-59.
- Park, Y.H., J.G. Kim, Y.W. Shin, H.S. Kim, Y.J. Kim, T. Chun and K.Y. Whang, 2008. Effects of *Lactobacillus* acidophilus 43121 and a mixture of *Lactobacillus* casei and Bifidobacterium longum on the serum cholesterol level and fecal sterol excretion in hypercholesterolemiainduced pigs. Biosc. Biotechnol. Biochem., 72: 595-600.
- Piao, X. S., I. K. Han, J. H. Kim, W. T. Cho, Y. H. Kim, and C. Liang. 1999.
 Effects of Kemzyme, Phytase, and Yeast Supplementation on the Growth Performance and Pullution Reduction of Broiler Chicks. *Asian-Aust. J.Anim.Sci.* 12 (1): 36-41.
- Putra, A.N., N.B.P. Utomo and Widanarni. 2015. Growth Performance of Tilapia (*Oreochromis niloticus*) Fed with Probiotic, Prebiotic and Synbiotic in Diet. Pakistan Journal of Nutrition 14 (5): 263-268

- Queiroz, R.C., A.F. Bergamaschine, J.F.P. Bastos, P.C. Santos and G.C. Lemos, 2004. Uso de produto a base de enzima e levedura na dieta de bovines: Digestibilidade dos nutrients e desempenho em confinamento. Rev. Brasil Zootech., 33: 1548-1556.
- Roni, N. G. K., E. Puspani, Dan I G. N. G.
 Bidura. 2014. Efforts to Suppress Total Body Fat And Ammonia Gas excreta Ducks Management Through Feed Probiotics. The Magazine Scientific Ranch (Indonesia)18 (3):119-124
- Santin, E., A. Maiorka, M. Macari, M. Grecco, J.C. Sanchez, T.M.Okada and A.M. Myasaka, 2001. Performance and intestinal mucosa development of broiler chickens fed diets containing *Saccharomyces cerevisiae* cell wall. J. Applied Poult. Res., 10: 236-244.
- Santoso, U. and Sartini. 2001. Reduction of fat accumulation in broiler chickens by *Sauropus androgynus* (Katuk) leaf meal supplementation. Asian-Aust. J. Anim. Sci. 14:346-350
- Siti, N.W., IGNG., Bidura, and I.A.P. The Utami. 2016. Effect of Supplementation Culture Cellulolytic Bacteria Isolated from the Rumen of Buffalo in the Tofu-Based Rations on the Performance and N-Nh3 Concentration in Excreta of Duck. J. Biol. Chem. Research. Vol. 33, No. 1. 214-225
- Stanley, V. G., R. Ojo, S. Woldesenbet, D. Hutchinson and L.F. Kubena. 1993.
 The Use of Saccharomyces sereviseae to Supress the Effects of Aflatoxicosis in Broiler Chicks. Poult. Sci. 72: 1867 1872.
- Steel, R.G. D. and J. H. Torrie. 1989.Principles and Procedures of Statistics.2nd Ed. McGraw-Hill International Book Co., London.
- Sutarpa, I. N. S., S. A. Lindawati, Y.

Ramona, I. N. S. Miwada, I. N. T. Ariana, and M. Hartawan, 2011. The Effect of Lactic Acid Bacteria Administration on The Performances, Total Bacteria in The Digestive Tract, and The Blood and Meat Cholesterol Content of Kampong Chickens. The 3rd International Conference on Biotechnology. Bioscience and Maintaining World Prosperity trhough Biotechnology Biosciences, and Revegetation. 21-22 September 2011. Udayana University, Denpasar Bali, Indonesia. Udayana University Press. Pp. 110-112

- Sudirman, I., 2004. Peranan Bakteri Asam Laktat dalam Kesehatan Hewan dan Peternakan, *Pelatihan Mikrobiologi Dasar*. Bogor: Fakultas Kedokteran Hewan IPB, Tanggal 26 April – 7 Mei 2004.
- Umiarti, A.T., E. Puspani dan IGNG. Bidura. 2014. Influence of *Saccharomyces spp* culture level in the ration on the performances and the levels of ammonia gas in chicken excreta. The Magazine Scientific Ranch (Indonesia) 17 (3): 79-84
- Utama, C. S. N. 2011. Potential of Probiotics rice bran. Poultry Indonesia.Vol VI, September: 78-80
- Wang, J. F., Y. H. Zhu, D. F. Li, H. Jorgensen, and B. B. Jensen. 2004. The influence of different fiber and starch types on nutrient balance and energy metabolism in growing pigs. *Asian-Aust. J. Anim. Sci. Vol.* 17 (2): 263-270
- Wu, Y., C. Lai, S. Qiao, L. Gong, W. Lu and D. Li. 2005. Properties of *Aspergillus Xylanase* and the Effects of Xylanase Supplementation In Wheat-Based Diets on Growth Performance and The Blood Biochemical Values In Broiler. Asian-Aust. J. Anim. Sci. Vol 18 (1): 66-74

- Yamada, E.A. and V.C. Sgarbieri, 2005. Yeast (*Saccharomyces cerevisiae*) protein concentrate: Preparation, chemical composition and nutritional and functional properties. J. Agric. Food Chem., 53:3931-3936.
- Yi, Z., E. T. Kornegay and D. M. Denbow. 1996. Effect of microbial phytase on nitrogen and amino acid digestibility and nitrogen retention of turkey poults fed corn-soybean meal diets. *Poultry Sci.* 75: 979-990.
- Yousefi, M. and K. Karkoodi, 2007. Effect of probiotic thepax and *Saccharomyces cerevisiae* supplementation on performance and egg quality of laying hens. Int. J. Poult. Sci., 6: 52-54.