Food Science and Quality Management

ISSN 2224-6088 (print) ISSN 2225-0557 (online) Vol.43 2015

Quality

Management



performance

International Institute for Science, Technology & Education Accelerating Global Knowledge Creation and Sharing

About Journal of Food Science and Quality Management

Journal of Food Science and Quality Management is a peer reviewed journal published by HSTE. The journal publishes original papers at the forefront of food sciences, technologies and quality management topics. The journal is published in both printed and online versions.

Food Science and Quality Management is published by IISTE and follows a quarterly publication schedule. General inquiries and Paper submission: contact admin@iiste.org or FSQM@iiste.org

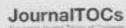
Index of this journal

















E z a Elektronische OO Zeitschriftenbibliothek











HSTE's acknowledgements to the supports from co-hosting universities worldwide

- University of North Carolina at Charlotte, United States
- California State University, United States
- · The City University of New York, United States
- · Aristotle University of Thessaloniki, Greece
- Universiteit Leiden, Netherlands

Vol 43 (2015) Table of Contents Articles Stabilizer Substance in Producing the Beet Juice Sati Yassin Ahmed Al-dalain 1-6 Comparison of Water Adsorption Characteristics of Plantain and Cocoyam in a Controlled Storage Condition. Abiodun A. Okunola, Joesph C. Igbeka, Caroline O. Okunola, John O. Ojediran. 7-13 Effect of Preservation Methods of Oil Palm (Elacisguincensis) Sap and Wine on the Mineral and Vitamin Compositions for Reproductive Health Theophilus M. Ikegwu, Jude O. Iwouno 14-20 Extraction of Phenolic Compounds from Wheat Bran using Ohmic Heating AsaadRehman Saeed Al-Hilphy, Ali Khudhair, laber AlRikabi, Abdullah M. Al-21 - 28Salim Influence of Selected Packaging Materials on Quality Aspects of High Pressure Processed Boccoli Puree Maryam Afrasiabi, YahyaMaghsoudlou, AlirezaSadeghiMahoonak, Mehran 29-35 Alami Socio - Economic Determinants of Fast Food Industry Growth in Pakistan AbdullGafoorAwaan Muhammad Nawaz 36-47 Physico-Chemical and Functional Properties of Cookies Produced from Sweet Potato- Maize Flour Blends Adeyeye, Samuel A., Akingbala, John O. 48-57 Prevalence and Growth Potential of Staphylococcus Aureus and Salmonella Spp. Isolated from Paper Currencies, Jimma Town, Southwest Ethiopia GosaGirma 58-72 The Creation of Barracuda Fish Based Meatball as Nutritious Food Alexander Hariohoedojo, Hari Minantyo, PrasetyonSepsiWinarno 73-78 Germination Effect on Functional Properties and Antitrypsin Activities of Pigeon Pea (Cajanuscajan (L.) Millsp.) Sprout Flour Ni WayanWisaniyasa, 1 Ketut Suter, YustinusMarsono, IN. Kencana 79-83 Fasciolosis: Prevalence, Evaluation of Flotation and Simple Sedimentation Diagnostic Techniques and Monetary Loss due to Liver Condemnation in Cattle Slaughtered at WolaitaSoddo Municipal Abattoir, Southern Ethiopia AssefuAlemu Belay Abebe 84-97 Effect of Fermentation on the Anti-Nutritional Factors and D. C. Okafor, A. I. Peter-Ikechukwu, R. O. Enwereuzoh, A.E. Uzoukwu, S.

M. Nze, M. I. Agunwa, F. I. Anagwu, C. Onyemachi

98-111

Kecambah Kacang Gude

by Wayan Wisaniyasa

FILE ARTIKEL_FINAL_PUBLIKASI_INTERNAS_KEC_KAC_GUDE.DOCX

(45.95K)

 TIME SUBMITTED
 04-FEB-2016 11:46PM
 WORD COUNT
 3580

 SUBMISSION ID
 627792547
 CHARACTER COUNT
 20011

GERMINATION EFFECT ON FUNCTIONAL PROPERTIES AND ANTITRYPSIN ACTIVITIES OF PIGEON PEA (Cajanuscajan (L.) Millsp.)SPROUT FLOUR

¹Wisaniyasa, N.W., ¹Suter, I.K., ²Marsono, Y., and ¹Putra, I.N.K.

¹Departement of Food Science and Technology, Udayana University, Bukit Jimbaran, Badung, Bali, Indonesia

²Departement of Food and Agricultural Product Technology, GadjahMada University, Yogyakarta, Indonesia

Email: wisaniyasa_2007@yahoo.com

Abstract: Germination is a series of changes in morpholog 1 physiology and biochemistry. These changes are likely to affect the nutritional content, anti-nutritional and functional properties of sprout flour produced. The purpose of this study was to determine the effect the length of germination on the functional properties and the antitrypsin activity of pigeon pea sprout flour. After being soaked at 50 °C during the first 6 hours, followed by soaking at room temperature for 30 hours, pigeon pea seeds were germinated at room temperature with germination time of 24; 36, 48; 60 and 72 hours respectively. The resulting sprouts were dig 37 at 50 °C, then powdered. The results showed that the germination process increased water absorption capacity and the oil absorption of the pigeon pea 20 uts flour 15.73% and 14.80% respectively. The longer the time of germination, the higher the water absorption capacity and the oil absorption were. The process of germination decreased pigeon pea flour swelling volume from 5.3% to 4.2% in pigeon pea sprouts flour, but increased the solubility of 13.3% (pigeon pea flour) to 19.0% (pigeon pea sprouts flour) and speeding wettability from 377 seconds (pigeon pea flour) to 188 seconds (pigeon pea sprouts flour). The longer the time of germination, the activity of trypsin inhibitor decreases. At the 72-hour germination, anti-trypsin activity decline reached 46%, from 573.090UTI / g sample becoming 308.827 UTI /g sample.

Keywords: germination, functional properties, antitrypsin, pigeon pea

INTRODUCTION

Pigeon pea beans contain complete nutrients that are not inferior to soy. The contents of nutrients of pigeon pea in 100 g is energy 336 cal., protein 20.7 g, fat 1.4 g, carbohydrates 62.0 g, 125 mg calcium, phosphorus 275 mg, 4.0 mg iron, vitamin A 19RE and vitamin B1 0.48 mg (Odeny, 2007). Until now, most of the pigeon pea has been used as a vegetable. Like other legumes, pigeon pea can also be processed into tempeh, concentrate, isolate protein and

pigeon pea flour (Odeny, 2007). Among the processed products, pigeon pea flour has relatively good prospects.

One of the obstacles of the use of bean flour is the presence of anti-nutritional substances such as anti-trypsin which may decrease the bioavailability of nutrients in it. One of the efforts to overcome this is to give pretreatment before flouring to eliminate anti-nutritional substances, namely by means of germination (Sharma and Shegal, 1992; Savelkoulet al., 1994; Adawyet al., 2003; Ramakrishna et al., 2006; Valdez-Anguino, 2015)

Germination has been known as a process that is not expensive and is technologically-effective in improving the nutritional quality of beans (Steve, 2012 and Valdez-Anguino, 2015). Duenaset al. (2009) proved that germination is able to increase the nutrient content, digestibility and availability of free amino acids, dietary fiber and bioactive components of lupine seeds (Lupinusangustifolius L.). Adawyet al., (2003) found that the germination green beans, peas and lentile proven to increase mineral content of N, K, Ca, P, Mg, Fe and Mn. Germination can cause changes in the functional properties (Nowzuet al., 2010), chemical composition (El-Moneimet al., 2012) as well as the content of antinutritional substances such as anti-trypsin (Savelkoul, 1994). No research has been done on the long period effects of germination on the functional nature and content of the flour anti-trypsin of the pigeon pea sprouts produced. Therefore, this research is very necessary. The purpose of this study was to determine the effect of length germination time on the functional properties (water absorption capacity, oil absorption capacity, swelling volume, solubility and wettability) and anti-trypsin activity of the pigeon pea sprout flour produced.

MATERIALS AND METHODS

Materials used in this study were harvest dried pigeon pea from Kubu Village, Kubu District, Karangasem Regency. Materials for chemical analysis used were N-α-benzoyl-L-arginine-4 nitroanilide hydrochloride (BAPNA), NaOH, a solution of acetic acid, trypsin solution (Tripzin-1x-S from porcine pancreas, Sigma-Aldrich), BAPNA solution. The tools used included UV / Vis spectrophotometer to analyze the levels of anti-trypsin.

This research used completely randomized design, with a length of germination time namely 24, 36, 48, 60 and 72 hours with three replications. The

research was preceded by the making of pigeon pea sprouts. The procedure of making pigeon pea sprouts was done in the following manner: pigeon pea was sorted and then soaked at 50 °C in the first 6 hours, then at room temperature for the next 30 hours. Replacement of water was done every 6 ours, then drained, and germinated in a leaf-base plastic basket container. The length of germination time was done according to treatment. Pigeon pea amount in each treatment was 150 g with a thickness of 1 cm and was incubated at room temperature. Each treatment was evenly sprinkled with water of 10 ml every 12 hours. Having obtained the pigeon pea sprouts, then dried in an oven at 50 °C for 8 hours, then ground and sifted using a sieve of 60 meshes, in order to obtain pigeon pea sprouts flour. Three repetitions of the experiment were done. The observed parameters were the functional properties and anti-trypsin activity of pigeon pea flour sprouts. Analysis of functional properties include water absorption capacity by gravimetric method (Fernandez-Lopez et al., 2009), oil absorption capacity gravimetric method (Fernandez-Lopez et al., 2009), swelling volume with volumetric method (Collado&Corke, 1999), solubility with gravimetric method

(Collado&Corke, 1999), and wettability with hydration methods (Bhandari, 2000). Analysis of antitrypsin activity was made by means of enzymatic methods (Kakadeet al., 1974 within Pazlopez, 2012). Data were analyzed using SPSS, when there was an effect of treatment; the analysis was continued with Duncant test (Steel and Torrie, 1995).

RESULTS AND DISCUSSION

The influence of the length of time of germination on water absorption capacity, oil absorption capacity, swelling volume, solubility and wettability of pigeon pea flour sprouts can be seen in Table 1.

Table 1. The water absorption capacity, oil absorption capacity, swelling volume, solubility and wettability of pigeon pea sproutsflour at various lengths of germination time

Length of germination (hour)	Water absorption capacity (% db)	Oil absorption capacity (% db)	Swelling volume (% db)	Solubility (% db)	Wettability (second)
0	216.65	173.96	5.3	13.32	377
	± 1.64 ^d	± 1.64 ^d	± 0.10 ^a	± 0.91 ^b	± 13 ^a
24	224.32	181.27	5.10	13.73	326
	± 2.73 °	± 2.87 °	± 0.10 ^a	±1.22 b	±8 b
36	237.71	182.71	4.77	14.34	275
	± 1.09 ^b	±2.26 °	± 0.06 ^b	± 1.98 ^b	± 10 °
48	248.56	184.16	4.73	16.95	214
	± 1.09 ^a	± 2.73 °	± 0.12 b	± 1.62b ^a	±2 b
60	250.01 ±1.66 ^a	192.48 ± 2.73 ^b	$^{4.57}_{\pm0.06}$ b	18.08 ± 1.39 ^a	204 ± 4 ^b
72	250.73	199.72	4.23	19.05	188
	± 1.09 ^a	± 3.91 ^a	± 0.21 °	± 1.04 ^a	± 2 ^a

a,b,c, Value in a column with unlike superscript letters were significantly different (p<0.01)

Water Absorption Capacity

Table 1 shows that the germination process can increase water absorption capacity of pigeon pea sprouts flour. The smallest water absorption capacity was 224.32% (db) obtained from the pigeon pea sprouts flour with germination time of 24 hours, while the largest water absorption capacity was 250.73% (db) obtained from pigeon pea sprouts flour with long germination period of 72 hours but was not significantly different from pigeon pea sprouts flour with germination time of 48 hours and 60 hours. Increased water absorption capacity was probably due to the

fact that the process of germination can increase levels of dietary fiber of the germinated pigeon pea seed. This is in line with the chemical composition of pigeon sprouts flour in this research. It was shown that germination for 48 hours increased the dietary fiber content from 16.85% (pigeon pea seed) to from wheat flour germinated. This is in line with the chemical composition of pigeon pea sprouts flour in this research. It was shown that germination for 48 hours increase the dietary fiber content from 16.85% (pigeon pea seed) to 18.46% (pigeon pea sprouts). It was reported that one of the important dietary fiber

properties is high water binding ability 1995). Increased (Marsono, water absorption capacity may also be due to increased levels of protein and quality of protein so that absorption capabilities and increased interaction with water (Chauhan and Sing, 2013). This study also found that pigeon pea flour protein content was 23.12%, while the protein content flour pigeon pea sprouts with germination time of 48 hours was 25.32%. Previous researcher (Adedejiet al., 2014) reported germination increases water absorption capacity of corn starch. Similar trend was found in sprouts amaranth flour (Chauhan and Sing, 2013) and tigernut sprouts flour (Chinmaet al., 2009).

Oil Absorption Capacity

Germination was also able to increase the oil absorption capacity of pigeon pea flour bean sprouts. The smallest oil absorption capacity was 181.27% (db) obtained from the flour pigeon pea sprouts with germination time of 24 hours, but it was not statistically significantly different from germination time of 36 and 48 hours. Largest oil absorption capacity was 199.72% (db) obtained from pigeon pea sprouts flour with the germination time of 72 hours. Increased oil absorption capacity of pigeon

pea sprouts flour can be caused by a decrease in fat content. This study found that the germination for 48 hours was able to lower the fat content of 4.9%. It is likely due to decreased levels of fat because fat is used as an energy source for the growth of sprouts. So the longer the germination, the more oil can be tied so that the oil absorption capacity is increasing. Similar results were reported by Adedejiet al. (2014) who found that the germination increase oil absorption capacity of cornstarch. A similar case was reported by Chinmaet al (2009) who proves that the germination can increase the absorption capacity of the tiger nut flour oil.

Swelling volume

Germination reduces the ability swelling volume of pigeon pea sprouts flour. The smallest swelling volume 4.23% (db) was obtained from pigeon pea sprouts flour with germination period of 72 hours; while the largest swelling volume was 5.10 (db) obtained from the pigeon pea sprouts flour with germination time of 24 hours. The decreased capability of flour swelling volume was possibly due to the fact that, during germination, starch was hydrolysed to simpler compounds. The bond of α 1-4 in a starch was disrupt by amylase and

subsequently by α-glycosidase. This reaction caused the starch to change into simpler sugar (Egwim and Ademonom, 2009). The decreased level of starch possibly caused swelling volume to decrease. Adedejiet al. (2014) also found the same thing, that is, germination decreased the ability of cornstarch swelling power. In the nongerminated corn flour, swelling power capability was 19.81 mg/g, while in the flour of the corn germinated for 24, 48 and 72 hours, the swelling power became 19.1 mg/ g, 18.8 mg/g, and 15.0 mg/g respectively. Elhkalifa and Bernhard (2013) also reported that germination of sorghum flour caused swelling power to decrease. Swelling power of sorghum which was not germinated was 6.3 g/g, while sorghum which was germinated for 24, 48 and 72 hours was 5.5 g/g, 5.2 g/g and 4.9 g/g respectively.

Solubility

Germination increases the solubility of pigeon pea sprout flour. The smallest solubility of 13.73 % (db) was obtained from pigeon pea sprout flour with germination of 24 hours but not significantly different from the germination of 36 hours, while the largest solubility of 19.05% (db) was obtained from bean sprouts pigeon pea flour with germination period of 72 hours,

significantly different but not germination of 48 and 60 hours. Increased solubility of starch as a result of germination is likely due to the fact that during the germination process there was hydrolysis from complex compounds into simpler compounds that are more water soluble and increased the activity of several enzymes such as amylase and protease enzymes. Enzymes involved in the hydrolysis of food reserves were α-amylase, β-amylase and According protease. to Phattanakulkaewnorie et al.(2011),increased amylase enzyme activity was due to germination causing more and more complex molecules such as carbohydrates break down into sugar molecules that are simpler and thus easier to dissolve.

Results of this study were supported by the research of Hussein *et al.*, 2013, which proves that the germination can improve the solubility of the starch of mung bean sprouts. Other studies have also proved that the solubility of the germinated sorghum flour increased from 14-50%, in line with the increase in temperature of 55-95°C, whereas the solubility of nongerminated sorghum flour was lower that is, the increase of 10-24% was in line with increases in temperature 55-95 °C (Phattanakulkemorie*et al.*, 2011). Research

by Atifyet al. (2012) also proved that the germination can increase the protein solubility of sorghum flour.

Wettability

Table 1 shows that the germination is capable of lowering wettability of pigeon pea flour sprouts. Wettability is the time required by the flour to absorb water or the time required by the flour to wet. The fastest wettability was obtained from flour 188.00 seconds pigeon pea sprouts with the germination time of 72 hours, while the longest was 326.00 seconds wettability obtained from flour pigeon pea sprouts with germination time of 24 hours. This is supported by research of Nwosuet al. (2010), which examines the effect of the germination on wettability of the asparagus seed flour. They found that the seeds of asparagus which have not been germinated had wettability of 189 minutes, while those germinated for 72 hours had the wettability of 79 minutes. The other research by Nwosu (2010), namely on seed germination of Oze also proved that the germination lowered wettability of Ozu flour. The study proves that the wettability of Ozu flour seed not germinated was 5 minutes, while Ozu seeds germinated for 24 hours were 4 minutes. The reduced wettability of flour was likely

caused by germination which was able to increase hydrolysis of complex compounds into simpler compounds. The formation of compounds which were simpler due to the increasing of the amount of hydrophilic compounds resulted during germination, so that the time required to be wet was increasingly short.

Trypsin inhibitor activity

Effect of germination time on the activity of trypsin inhibitor can be seen in Table 2. Pigeon pea beans that have not been germinated had trypsin inhibitor activity of 573.090 UTI / g sample. From Table 2 it appears that the longer the time of germination, the activity of trypsin inhibitor decreases.

Table 2. Activities of trypsin inhibitor of pigeon pea sprouts flour (UTI)

Activity of trypsin inhibitor (UTI)/g sample	Decrease of anti-trypsin activity after germination (%)
573.090 ± 37.903	0
406.923± 32.835 ^a	29±3.08 ^a
376.742± 13.400 ab	34±2. 38 ^{ab}
354.042 ± 8.785 ^b	38±4.88 b
348.754 ± 1.516 ^b	39±4.14 ^b
308.827 ± 24.216 °	46 ±2.27 °
	(UTI)/g sample 573.090 ± 37.903 406.923± 32.835 ^a 376.742± 13.400 ^{ab} 354.042 ± 8.785 ^b 348.754 ± 1.516 ^b

a,b,c, Value in a column with unlike superscript letters were significantly different (p<0.01)

Anti-trypsin is a protein compound that acts as anti-nutrients, that is, it has the ability to inhibit the activity of trypsin enzymes in the digestive tract. The mechanism of inhibition of trypsin enzyme activity by antitrypsin occurs due to the formation of complex bonds between the two substances. In this study, a decrease in antitrypsin activity as a result of germination was possible because during germination process hydrolysis reaction took place. In the process, there was also a proteolysis by a more active endogenous protease enzyme. The presence of endogenous protease enzyme can hydrolyze antitrypsin during development of sprouts to be used as a source of sulphur amino acids (Valdez-Anguinoet al., 2015). This is in line with the research results of Shegal and Sharma (1992), which proved that faba seed

germination for 2 days can lower the antitrypsin activity by 65%. Friaset al. (1995) reported that the lentil seed germination for 72 hours was able to lower antitrypsin activity by 10%. Seed germination of green beans, peas and lentils for 72 hours was able to significantly decrease the antitrypsin activity, but if germination extended to 120 hours, antitrypsin activity actually increased (Adawyet al., 2003). Other studies have reported that the germination of Indian bean (Delicos lablab) for 32 hours was able to lower antitrypsin activity by 17% (Ramakhrisnaet al., 2006). Malomoet al., (2012) reported that soybean germination for 72 hours was able to lower flour antitrypsin activity of 29.82%

CONCLUSION

Germination process increased absorption capacity, water absorption capacity and solubility of pigeon pea sprouts flour. On the other hand, the process of germination decreased swelling volume, wettability value and anti-trypsin activity of pigeon pea sprouts flour. percentage decrease in antitrypsin activity on the germination of 24-72 hours ranged from 29 to 46%.

25 ACKNOWLEDGMENTS

The author would like to thank
the Rector of Udayana University for
financial support given to me during
my studies at the Doctoral Study
Program of Agricultural Sciences
Udayana University.

REFERENCES

Adawy, TAE.,Rahma E.H., Bedawey,
A.A.E. and Beltagy, A.E.E. 2003.
Nutritional Potential and
Functional Properties of
Germinated Mung bean, Pea and
Lentil Seeds.Journal Plant Foods
for Human Nutrition. 58 (3):1-

3

Adedeji, OE., Oyinloye, O.D. and Ocheme,
O.B. 2014. Effects of Germination
Time on The Functional Properties
of Maize Flour and the Degre of
Gelatinization of Its
Cookies. African Journal of Food
science. 8(1): 42-47.

- Afify, A.E.M., El-BeltagiH.S., El-Salam, S..MA.andOmran, A.A. 2012 Solubility, Digestibility and Fractionation after Germination of Sorghum Varieties. Downloaded from:journals.plos.org/plosone/article?id=10.1371/journal.pone.0031
- Bhandari, B. 2000. Understanding
 Food: Priciples and
 Preparation. Wadsworth
 Thomson Learning. USA.
 Downloaded from :www:
 Fst.edu.av/staff/bbhandari/teac
 hing/brown.Amy.
- Chauha 11 A., and Singh, S. 2013.
 Influence of Germination on Physico chemical Properties of Amaran 11 (Amaranthus Spp.)
 Flour. International Journal of Agriculture and Food Science Technology. ISSN 2249-3050. 4 (3): 215-220.
- Chinma, C.E., Adewuyi, O.andAbu, J.O. 2009.Effect of Germination on the Chemical, Functional and Pasting Properties of Flour from Brown and Yellow Varieties of Tigernut (Cyperusesculentus).Food Research International 42.p.1004–1009.
- Collado, L.S., and Corke, H. 1999. Heat
 Moisture Treatment Effect on
 Sweet Potato Starches Differing in
 Amylosa Content. Journal of Food
 Chemistry. 65: 339-346.
- Dueñas, M., Hernández, T., Estrella, I., &Fernández, D.. 2009.

 Germination as a Process to Increase the Polyphenol Content and Antioxidant Activity of Lupin Seeds (Lupinusangustifolius L.).

Journal of Food Chemistry, 117(4): 599-607.

Egwim E. and Ademonom. 2009.

Predicting α-amylase Yield and
Malt Quality of Some sprouting
Cereals using 2nd Order
Polynomial Model. African
Journal Biochemistry 3 (8): 288-

El-Moneim A., Afify, M.R. El-Beltagi,
H.S., Samiha, M.A., El-Salam
and Omran, A.A. 2012. Protein
Solubility, Digestibility and
Fractionation after Germination of
Sorghund Varieties. Research
Article. Editor: Vladimir N.
Uversky, University of South
Florida College of Medicine,
United States 27 Imerica. PLoS
ONE 7(2): e31154.
doi:10.1371/journal.pone.0031154

Fernandez-Lopez, Sendra-NatalJ.,

Varro, E.,Sayas, C., Martos,
E.V. and Perez-Alvarez. 2009.

Storage Stability of a High Dietary
Fibre Powder From Orange byProducts. International Journal
Food Science and Technology
44:748-756.

Frias J., Diaz-Pollan, C., Hedley, C.L. and Vidal-Valvedley, C.. 1995.
Evolution of Trypsin Inhibitor Activity During Germination of Lentils. Journal Agricultural Food Chemistry.,43(8): 2231–2234

Hussain, M.S., Anjum, Unin, M.B., and Ali, S.. 2013. Optimization of Germination Effect on Functional Properties of Mungbean Flour By Response Surface Methodology. Pakistan Journal of Science 65 (2):214-218.

Malomo O., Ogunmoyela, O.A.,
wajoba, B.S.O., and Kukoyi, I.
Effect of Germinated Soy Flour on
The Sensory Acceptability of SoyWheat Composite Bread Asian
Journal of Natural and Applied
Sciences

Marsono, Y. 1995. Fermentation of Dietary Fibre in the Human Large Intestine: A review. Indonesian Food and Nutrition Progress, 2: 48-53.

Nwosu, J.N. 2010. The Effects of Processing on Functional the Properties of 'Oze' (Bosqueiaangolensis) Seeds. Pakistan Jou 24 of Nutrition 9 (8): 781-786. 2010 ISSN 5194 © Asian Network for Scientific Information

Nwosu, J.N., Ogueke, C.C., Owuamanam,
C.I. and Iwouno, J.O.. 2010.
Functional Properties and
Proximate Composition of
Asparagus Bean (Vigna
Sesquipedalis) as Influenced by
Malting. Journal of American
Science 6(9), p 376-82

Odeny, D.A. 2007. The potential of Pigeonpea (Cajanuscajan (L.) Millsp.) in Africa. Natural Resources Forum.Blackwell Publishing Ltd 31. 297–305.

Pazlopez, C.M. 2012. Common Beans
Cooked at High Altitudes Have
Higher Trypsin Inhibitor Activity
and Lower Protein Digestibility
Than Beans Cookies at Sea Level.
Presented to The College of
agriculture and Life Science,
Physical Sciences, Cornel
University, America.

Phattanakulkaewnorie, Paseephol, T., andMoongngarm, A. 2011.
Chemical Compositions and Physico-chemical Properties of Malted Sorghum Flour and Characteristics of Gluten Free Bread.Word Academy of Science, Engineering and Technology. Vol.5.

Ramakrishna, Rani, J.andRao, P.R.. 2006.

Anti-Nutritional Factors During
Germination in Indian Bean
(Dolichos lablab L.) Seeds. World
Journal of Dairy & Food Sciences
1 (1): 06-11.ISSN 1817-308X ©
IDOSI Publications

Savelkoul, F.H., Tamminga, S.,
Leenaars,P.P., Schering, J. and
Maaat, D.W.T. 1994.
TheDegradation of Lectins,
Phaseolin and Trypsin Inhibitors
During Germination of White
Kidney Beans (*Phaseolus vulgaris*L).Plant Foods Hum Nutr. 45(3), p.
213-22.

Sharma and Shegal. 1992. Effect of
Domestic Processing, Cooking
and Germination on the Trypsin
Inhibitor Activity and Tannin
Content of Faba bean (Viciafaba).
Plant Foods
HumanNutrition42(2):127-33.

Steve, I.O. 2012. Influence of Germination and Fermentation on Chemical Composition, Protein Quality and Physical Properties of Wheat Flour (*Triticumaestivum*). Journal of Cereals and Oil Seeds 3(3): 35-47

Valdez-Anguino, D.M., Herrera-

Cabrera, E., Davila-Ortiz, G., OOMAH, B.D., Cardador-Martinez, A., and Jimenez-Martinez.C. 2015. Content and distribution of Nutritional and 321-Nutritional Compounds During Germination of Three Mexican Faba Bean (Viciafaba). International Journal of Research In Agriculture and Food Sciences. Vol. 2, No.9 ISSN 2311 -2476

Kecambah Kacang Gude

ORIGINALITY REPORT

19% SIMILARITY INDEX

13%

15%

11%

LARITY INDEX INTERNET SOURCES

PUBLICATIONS

STUDENT PAPERS

PRIMARY SOURCES

Adegunwa, M.O., A.A. Adebowale, H.A. Bakare, and K.K. Kalejaiye. "Effects of Treatments on the Antinutritional Factors and Functional Properties of Bambara Groundnut (V oandzeia subterranea) Flour: Bambara Flour", Journal of Food Processing and Preservation, 2013.

Publication

2 Submitted to Udayana University
Student Paper

2%

O., Ocheme B., Adedeji O. E., Lawal G., and Zakari U. M.. "Effect of Germination on Functional Properties and Degree of Starch Gelatinization of Sorghum Flour", Journal of Food Research, 2015.

2%

Publication

Submitted to Higher Education Commission Pakistan

1%

Student Paper

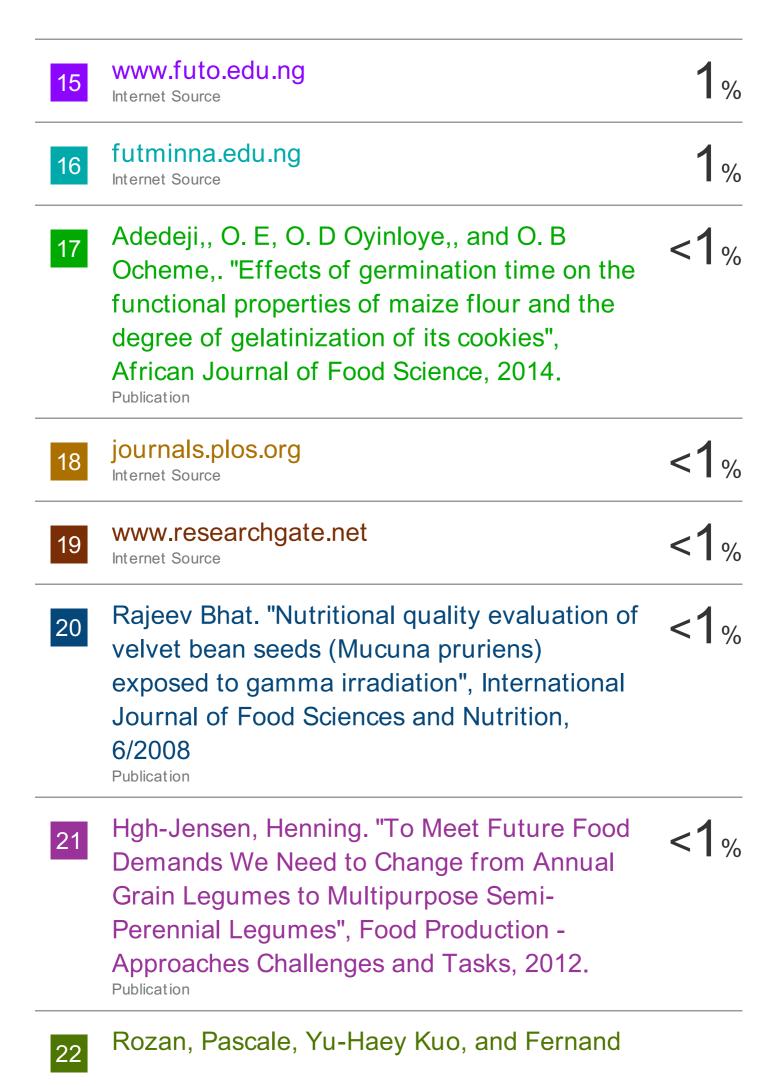
5

soeagra.com

Internet Source

1%

6	I. Rotari. "Calcium Ions Make Phytohemagglutinin Resistant to Trypsin Proteolysis", Journal of Agricultural and Food Chemistry, 2008.	1%
7	Ana Vale. "Valorização de germinados de Brassica oleracea através da avaliação nutricional e da composição em compostos bioactivos", Repositório Aberto da Universidade do Porto, 2015. Publication	1%
8	www.scihub.org Internet Source	1%
9	www.lablablab.org Internet Source	1%
10	file.scirp.org Internet Source	1%
11	www.ripublication.com Internet Source	1%
12	mgb.ugm.ac.id Internet Source	1%
13	Hagenimana, Anastase, and Xiaolin Ding. "A Comparative Study on Pasting and Hydration Properties of Native Rice Starches and Their Mixtures", Cereal Chemistry, 2005. Publication	1%
4.4	www.bioline.org.br	1



Consumers", Journal of Agricultural and Food Chemistry, 2000. Publication www.savap.org.pk <1% 23 Internet Source www.pjbs.org 24 Internet Source Gurmeric, Vildan Er, Mahmut Dogan, Omer 25 Said Toker, Ercan Senyigit, and Nevruz Berna Ersoz. "Application of Different Multicriteria Decision Techniques to Determine Optimum Flavour of Prebiotic Pudding Based on Sensory Analyses", Food and Bioprocess Technology, 2013. Publication www.insinet.net <1% Internet Source Afify, Abd El-Moneim M. R., Hossam S. El-Beltagi, Samiha M. Abd El-Salam, and Azza A. Omran. "Protein Solubility, Digestibility and Fractionation after Germination of Sorghum Varieties", PLoS ONE, 2012. Publication www.academicjournals.org 28

Internet Source

Lambein. "Free Amino Acids Present in

Commercially Available Seedlings Sold for

Human Consumption. A Potential Hazard for

<1%

Ahuja, Heena, Satvir Kaur, Anil Kumar Gupta, Sarvjeet Singh, and Jagmeet Kaur. "Biochemical mapping of lentil (Lens culinaris Medik) genotypes for quality traits", Acta Physiologiae Plantarum, 2015.

<1%

Publication

Nkundabombi, Marie Grace, Dororthy Nakimbugwe, and John H. Muyonga. "Effect of processing methods on nutritional, sensory, and physicochemical characteristics of biofortified bean flour", Food Science & Nutrition, 2015.

<1%

Publication

Singh, Arashdeep, and Savita Sharma.
"Bioactive Components and Functional
Properties of Biologically Activated Cereal
Grains: A Bibliographic Review", Critical
Reviews in Food Science and Nutrition, 2015.

<1%

Publication

Shimelis, E.A.. "Effect of processing on antinutrients and in vitro protein digestibility of kidney bean (Phaseolus vulgaris L.) varieties grown in East Africa", Food Chemistry, 2007

<1%

Publication