watermelon

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IDENTIFICATION OF MICROBES ANTAGONISTIC AGAINST Fusarium oxysporum ISOLATED FROM RHIZOSPHERE ZONE OF WATERMELON

IDENTIFIKASI MIKROBA ANTAGONIS Fusarium oxysporum YANG DIISOLASI DARI RHIZOSFER TANAMAN SEMANGKA

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ABSTRACT

This research was aimed to isolate and identify microbes which antagon 15 against *Fusarium oxysporum*, the causative agent of vascular wilt in watermelon plants. The antagonistic microbes were isolated from soil samples collected from rhizosphere of Gatermelon farm located at west Sanur village, South Denpasar, Bali. Isolation of fungi and bacteria were conducted on potato dextrose agar medium (PDA) and nutrient agar medium (NA), respectively. Fungal isolates were then observed under light microscope for its morphological characteristics before identification using a reference book. Bacterial isolates were characterized using various tests, such as gram stain reaction, existence of endospores, catalase reaction, a billity to ferment various sugars. Their characteristics were then compared with those described in a reference book. Two fungal species (*Trichoderma harzianum* and *Trichoderma viride*) and two bacterial for the growth of *F. oxysporum*.

Keywords: Bacillus sp., Pseudomonas sp., Trichoderma harzianum, Trichoderma viride, Bali, Watermelon

INTISARI

Penelitian ini buzujuan untuk mengisolasi dan mengidentifikasi mikroba yang bersifat antagonis terhadap *Fusarium* oxysporum yaitu penyebab penyakit layu vaskular pada tanaman semangka. Mikroba antagonis diisolasi dari sampel tanah yang diambil dari rhizospher tanaman semangka yang ditanam di perkebunan yang berlokasi di desa Sanur Barat, Denpasar Selatan, Bali. Isolasi fungi dilakukan pada dextrose agar medium (PDA) dan isolasi bakteri dilakukan pada nutrient agar medium (NA). Isolat fungi diamati dengan microskop cahaya untuk mengetahui ciri-ciri morfologi sebelum diidentifikasi menggunakan buku reference. Isolat bakteri dikarakterisasi menggunakan beberapa uji, meliputi, reaksi pewarnaan gram reaksi katalase dan kemampuan untuk melakukan fermentasi terhadap gula. karakteristik bakteri yang (Trichoderma harzianum dan Trichoderma viride) dan dua spesies bakteri (Pseudomonas sp. dan Bacillus sp.) yang berpotensi untuk dikembangkan menjadi agen biokontrol penghambat pertumbuhan F. oxysporum berhasil diisolasi dalam penelitian ini.

Kata kunci: Bacillus sp., Pseudomonas sp., Trichoderma harzianum, Trichoderma viride, Bali, Semangka

INTRODUCTION

According to report Indonesian Statistical Agent in 2011, watermelon production in Indonesia reached 474,327 tons in 2009. However, production of this fruit decreased sharply in 2010 where its production was only 348,631 tons. The main cause of this decrease was attributed by vascular wilt due to *Fusarium oxysporum* infection (Budiastuti *et al.*, 2012). This fungus is a *soil borne pathogen* causing many severe problems in many agricultural crops, including water melon. Plants infected by this fungus will show specific characteristics, such as stem rot with browny color and drying leaves. Until recently, watermelon farmers in west Sanur area rely only on the use of chemical-based fungicides to cope with this infection. Long term application of these fungicides has been claimed to have negative impact on ecosystems due to its long half-life (Djunaedi, 2009).

Based on the above background, a new and more environmentally friendly method, such as the use of natural enemy for this pathogen needs to be developed. Therefore in our research, microbes antagonistic against F.

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oxysporum were isolated from rhizosphere zone of watermelon farm located in West Sanur area. The main objective of this research was to obtain potential antagonist

MATERIALS AND METHODS

Sample collection

Fungal and bacterial antagonists were isolated from soils collected randomly from rhizosphere zone of watermelon farm in west Sanur area. Soil samples (100 g each) were aseptically collected from 6 locations in this area, placed in plastic bags, and transported to Microbiology Lab in School of Biology, Udayana University where isolation of microbes antagonistic against *Fusarium oxysporum* was conducted.

Isolation of antagonists

Fungal and bacterial antagonists were isolated by applying dilution and spread method as specified in Ramona and Line (2002), Ramona (2005) Suryanti *et al.* (2013), and Wulansari *et al.* (2015). Samples were diluted up to dilution rate of 10⁻⁶ and spread on potato dextrose agar medium (PDA) for fungal isolation or on nutrien agar medium (NA) for bacterial isolation. All inoculated media were incubated at ambient temperature for 2-5 days until bacterial or fungal colonies grew. Colonies that showed antagonistic activities against *F. oxysporum* in *in vitro* dual culture assays were isolated and identified at least up to genus level.

Identification of bacterial and fungal antagonists

Fungi and bacteria that showed antagonistic activities against F. oxysporum in the in vitro assays were identified up to genus or species level when necessary based on their morphological and physiological characteristics. Fungal isolates were observed under a light microscope for their hypha morphology, conidiophore, their spore structures, and colour on PDA. Their characteristics were then compared with that specified in a reference book of Fungi and Food Spoilage (Pitt and Hocking, 1997). For bacterial identification, their characteristics observed included gram stain reaction, endospore existence, catalase reaction, ability to ferment various sugars (glucose, lactose, maltose and sucrose), reaction on indol, and motility on SIM medium. The characteristics of each bacterial isolate were zen compared with that specied in reference book of Bergey's Manual of Determinative Bacteriology 9th Edition (Holt et al., 1994).

RESULTS

Two potential fungal and two bacterial antagonists were successfully isolated in this project. The microscopic and macroscopic (visualized with a Yazumi light microscope with 400 x magnification) characteristics of *T. Harzianum* P ISSN: 1410-5292, E ISSN: 2599-2856

isolates to be used to fight *F. oxysporum* infection in watermelon plants.

and *T. viride* at 5 days of aged on PDA medium are shown in Figure 1 and the microscopic characteristic of these two bacterial isolate is shown in Figure 2.

DISCUSSION

The two potential fungal antagonists were identified as Trichoderma harzianum and Trichoderma viride. The mycelium of the Trichoderma harzianum was initially white, and as a function of time it changed to green with grey pigmentation. Their Fialid grew on each branch tip where their clustered konidia with pale green were found. The size of their konidia, the length of their fialid, and the diameter of their hyphae ranged 2.63-3.07 µm, 7.28-8.54 µm, and 2.67-3.71 µm, respectively. Those characteristics matched with those specified in the identification reference book of Fungi and Food Spoilage (Pitt and Hocking, 1997). While the Trichoderma viride initially had pale green colonies and then changed into dark green. Their konidia are round and green, and located on the tips of their fialid. The size of their konidia, the length of their fialin, and the diameter of their hyphae ranged from 3.58 to 5.46 µm, from 7.62 to 8.46 µm, and from 2.60 to 2.74 µm, respectively. According to Pitt and Hocking (1997), both T. viride and T. harzianum are morphologically similar. The difference between them is the size of their konidia, where the size of T. viride konidia is bigger (ranging from 3.5 to $5.5 \,\mu$ m) than that of T. harzianum. The microscopic and macroscopic (visualized with a Yazumi light microscope with 400 x magnification) characteristics of T. harzianum and T. viride at 5 days of aged on PDA medium are shown in Figure 1.

T. harzianum and *T. viride* successfully isolated in this project were found to dominate the six locations of sampling points in the watermelon farm, in west Sanur area. This results is in line with that reported by Otadoh (2011) who stated that *Trichoderma* spp. are saprophytic fungi widely **3** read in soil samples. This type of fungi can grow rapidly in the rhizosphere zone of plants (Saba *et al.*, 2012). When grown on a mediun *in vitro*, their growth is characterizes by green konidia (Steyaert, 2007).

Fungi *Trichoderma* spp. has ability to prove cell wall degrading enzymes, such as cellulase (Raut *et al.*, 2014), chitinase (Sharma *et al.*, 2012), and glucanase (Saba *et al.*, 2012). These enzymes help their hyphae to penetrate lumen and assimilate the 7 cell wall of fungal pathogensFurthermore, Sharma *et al.* (2012), Hassan *et al.* (2013) and Benitez *et al.* (2004) reported that *Trichoderma* spp. also has the ability to produce antibiotics, such as harzianic acid, alamethicins, tricholin, peptaibols, alkil piron, massoilactone, viridin, gliovirin, glisoprenins, heptelidic acid, inhibiting the growth of other fungi around them.

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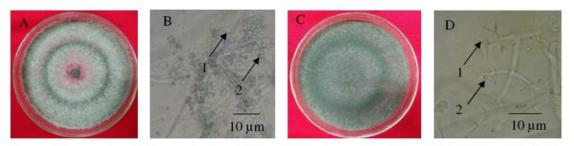


Figure 1. Macroscopic and microscopic characteristics of *T. harzianum* (A and B) and *T. viride* (C and D). Their Fialid and Konidia are shown by numbers 1 and 2, respectively.

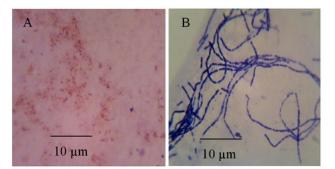


Figure 2. Microscopic characteristic of *Pseudomonas* sp. (A) and *Bacillus* sp. (B) observed under a light microscope (*Yazumi*) with magnification of 1000x

In addition to hydrolitic enzymes and antibiotics, some species of *Trichoderma* also produce *siderophores* for chelating Fe^{3+} ions in the soil so that the availability of these ions is limited for fungal pathogens (Benitez *et* **9**., 2004). Due to those characteristics, many species of *Trichoderma* spp. have been developed as biocontrol agents to control infections of *Fusarium* (Ojha and Chatterjee, 2012), *Rhizoctonia* (Seema and Devaki, 2012), and *Phytium* (Raut *et al.* 2014).

This research showed two bacteria 8 ntagonists were successfully isolated in this and they were identified as Pseudomonas sp. and Bacillus sp., based on the characteristics specified in the reference book of Bergey's Manual Determinative Bacteriology 9th Edition (Holt et al., 1994). Pseudomonas sp. isolate has greenish yellow pigment when grown on a solid medium in vitro. Other characteristics of this isolate observed in this research were Gram negative with cell size ranging from 0.86 to 1.91 x 0.2 to 0.67 µm, no endospore, catalase possitive, ferments sugars (glucose, maltose, and sucrose), motile on SIM medium, unable to ferment laktosa, and showed neative result for indol test. Whilst, Bacillus sp. isolate showed some characteristics, such as Gram possitive, chain cells with size of 2.44 to 2.90 x 0.61 to 1.73 µm, produced endospore, catalase possitive, fermented sugars

(glucose, maltose, and sucrose), motile on SIM medium, . negative indol test, and did not ferment lactose. The microscopic characteristic of these two isolate is shown in Figure 2.

11n the in vitro bioassays, these two bacterial antagonists inhibited the growth of Fusarium oxysporum (the causative agent of wilt in watermelon plant). The main cause of this inhibition was inconclusive and need to be further elucidated. Although it was inonclusive, some researchers reported that inhibition of *Pseudomonas* or *Dicillus* isolates may be due to sideropore production (Karimi et al., 2012), antibiosis (Sallam et al. 2013), competition (Jenifer et al., 2013), or other toxic compounds, such as HCN (Chaur and Lo, 1998). Ozaktan et al. (2015) and Vijayaraghavan and Vincet (2012) stated that antibiosis mechanism by Pseudomonas sp. and Bacillus sp. on fungal pathogens can happen through production of hydrolitic enzimes, such as cellulase, glucanase and chitinase. Toua et al. (2013) and Chaur and Lo (1998) reported that Pseudomonas sp. are also capable to produce antibiotics, such as phenazins, pyrrolnitrine and 2,4-diacetylphloroglucinol, in addition to the above hydrolitic enzymes.

JURNAL BIOLOGI UDAYANA22(2):71-74 CONCLUSION

Base on the morphological and physiological characteristics of 13 e isolates, the two fungal and bacterial isolates were identified as *T. harzianum*, *T. viride* and *Pseudomonas* sp. and *Bacillus* sp. Those microbes apper 10 d to be potential for bio control agent development to control the causative agent of wilt vascular disease in waterwatermelonmelon farm.

REFERENCES

- Badan Pusat Statistik. 2013. Produktivitas Tanaman Hortikultura Menurut Provinsi 2008-2011. BPS dan Dirjen. Hortikultura.
- Benitez, T., A. Rincon, M.C. Limon and A. Codon. 2004. Biocontrol mechanism of *Trichoderma* strains. Intl. J.Microbiology 7:249-260.
- Budiastuti, K., E.T. Tondok dan S. Wiyono. 2012. Causal agents of wilting Disease on Watermelon in Karawang, West Java. J. Fitopatologi Indonesia 8(4): 89-96.
- Chaur and T. Lo. 1998. General mechanisms of action of microbial biocontrol agents. Plant Pathology Bull. 7:155-166.
- Djunaedi, A. 2009. Biopestisida sebagai pengendali organisme pengganggu tanaman (OPT) yang ramah lingkungan. Embryo 6 (1):88-95.
- Hassan, R. Simon, Gowen, B. Pembroke. 2013. Use of *Trichoderma hamatum* for biocontrol of lentil vascular wilt disease: efficacy, mechanisms of interaction and future prospects. J. Plant Protection Res 53(1): 13-26.
- Holt, J.G., N.R. Krieg, P.H.A. Sneath, J.T. Staley and S.T. William. 1994. Bergey's Manual of Determinative Bacteriology 9th Edition. USA : Lippincott Williams and Wilkins.
- Jenifer, M. R. A., A. Reena, O.S. Aysha, S. Valli, P. Nirmala, P. Vinothkumar. 2013. Isolation of siderophore producing bacteria from rhizosphere soil and their antagonistic activity against selected fungal plant pathogens. Intl. J. Curr. Microbiology App. Sci. 2(1): 59-65.
- Karimi, K., J. Amini, B. Harighi, B. Bahramnejad. 2012. Evaluation of biocontrol potential of *Pseudomonas* and *Bacillus* Spp. against *Fusarium* wilt of chickpea. AJCS 6(4): 695-703.
- Ojha, S. and N.C. Chatterjee. 2012. Induction of resistance in tomato plants against *Fusarium oxysporum* f.sp. *Lycopersici* mediated through salicylic acid and *Trichoderma harzianumi*. J. Plant Protection Res. 52 (2): 220-225.
- Otadoh, J.A., A. Sheila, Okoth, J. Ochanda, J.P. Kahindi. 2011. Assessment of *Trichoderma* isolates for virulence efficacy on *Fusarium oxysporum* f.sp. *phaseoli*. Tropical and Subtropical Agroecosystems 13: 99-107.

P ISSN: 1410-5292, E ISSN: 2599-2856

- Ozaktan, H, Çakır, A. Gül, L. Yolageldi, A. Akköprü, Fakhraei, Akbaba. 2015. Isolation and evaluation of endophytic bacteria against *Fusarium Oxysporum* f. sp. *Cucumerinum* infecting cucumber plants. Austin J. Plant Bio. 1 (1): 1-6.
- Pitt, J.I. and A.D. Hocking. 1997. Fungi and food spoilage Vol. II, Ed. Pitt, J.I. and Hocking, A.D. London: Blackie Academic and Professional.
- Ramona, Y. and M.A. Line. 2002. Potential for large-scale production of a biocontrol fungus in raw and composted paper mill waste. Compost Science and Utilization. 10(1): 57-62.
- Ramona, Y. 2003. Assessment of some antagonist to fungal plant pathogens development of methods for their large scale cultivation. Ph.D.Thesis, The University of Tasmania. Australia.
- Raut, J., M. Calin, G. Vasilescu, M.B. Doni, T. Sesan, L. Jecu. 2014. Effect of non volatile compounds of *Trichoderma* Spp. against *Fusarium graminearum*, *Rhizoctonia* solani and *Pythium ultimum*. Scientific Bull.Series F. Biotechnologies 18: 178-181.
- Saba, Vibhash, Manisha, Prashant, Farhan, Tauseef. 2012. *Trichoderma* a promising plant growth stimulator and biocontrol agent. Mycosphere 3(4): 524-531.
- Sallam, N.A., S.N. Riad, M.S. Mohamed, A.S. El-eslam. 2013. Formulations of *Bacillus* Spp. and *Pseudomonas fluorescens* for biocontrol of cantaloupe root rot caused by *Fusarium Solani*. J. Plant Protection Res. 53(3): 295-300.
- Seema and Devaki. 2012. In vitro evaluation of biological control agents against *Rhizoctonia solani*. J. Agric.l Tech. 8 (1): 233-240.
- Sharma, Radheshyam, J. Arunabh and C.D. Ramesh. 2012. A brief review on mechanism of *Trichoderma* fungus use as biological control agents. Intl J. Innovations in Bio-Sciences 2 (4): 200-210.
- Steyaert, J.M. 2007. Studies on the regulation of conidiation in species of *Trichoderma*. (*Thesis*). Lincoln University.
- Suryanti, I.A.P., Y. Ramona, M.W. Proborini. 2013. Isolation and identification of the causative agents of wilting and their antagonistics in potato plants cultivated in Bedugul, Bali. J. Biologi 17(2): 37-41.
- Toua, D., M. Benchabane, F. Bensaid, R. Bakour. 2013. Evaluation of *Pseudomonas fluorescens* for the biocontrol of Fusarium wilt in tomato and flax. African J. Microbiology Res. 7(48): 5449-5458.
- Vijayaraghavan, P and S.G.P. Vincent. 2012. Purification and characterization of carboxymethyl cellulase from *Bacillus* sp. isolated from a paddy field. Polish J.Microbiology 61(1): 51–55.
- Wulansari, N.T. 2015. Upaya pengendalian penyebab penyakit busuk hitam pada tanaman brokoli (*Brassica oleracea* var. *italica*) dengan antagonisnya. (*Tesis*). Denpasar: Universitas Udayana.

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